



Biomonitoring Report
Menidia beryllina (EPA Method 1006) &
Americamysis bahia (EPA Method 1007)

prepared for
Corrosion Innovations
Client Contact: Jim Knocke

CHLOR*RID SP8 & CHLOR*RID SP8 Rinse
Run-off from Treated Plate Metal.

Non-protocol.

***M. beryllina*; EE USA Project No.: Q-625-13**

SURVIVAL 7-Day NOEC/LOEC = 25.0%/35.0% Lab Sample (LS)
GROWTH 7-Day NOEC/LOEC = <12.0%/12.03% LS
% CV = 6.4

***A. bahia*; EE USA Project No.: Q-626-13**

SURVIVAL 7-Day NOEC/LOEC = 1.1%/1.8% LS
GROWTH 7-Day NOEC/LOEC = 0.6%/1.1% LS
% CV = 14.9

Report Date: August 04, 2023
by

ENVIRONMENTAL ENTERPRISES USA, INC.
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This report contains seven pages plus seven appendices, A – G. This report must not be reproduced in part, only in whole. The results and conclusions presented in this report apply only to the sample(s) tested. All results should be considered valid unless otherwise noted in the report.

This is an amended report. The original report identified the product as “Corr-Ze 200 & Corr-Ze 100”. At the client’s request, the product name has been updated to CHLOR*RID SP8 & CHLOR*RID SP8 Rinse.

Michele Ellis
Michele Ellis
Effluents Testing Supervisor

David L. Daniel
David L. Daniel
President
QA/QC Officer

SWORN TO AND SUBSCRIBED BEFORE ME THIS
4 DAY OF August 20 23

Marie Betts #159677
Louisiana Notary Public Commissioned for Life
St. Tammany Parish * Statewide Jurisdiction

8/4/23
DATE

8/4/23
DATE



INLAND SILVERSIDE (*Menidia beryllina*) LARVAL SURVIVAL & GROWTH TEST
EPA-821-R-02-014: METHOD 1006

TEST OVERVIEW

A 7-day static-renewal toxicity test was conducted by Environmental Enterprises USA, Inc. (EE USA) to determine toxicity of LS CHLOR*RID SP8 & CHLOR*RID SP8 RINSE to *Menidia beryllina* larvae. Methods, materials, and results are presented in this document. Test organisms were cultured at EE USA and were 11-days-old when this test was initiated. Synthetic seawater was used as the laboratory performance control (LPC) solution and diluent in this test. Five replicates of the LPC solution and five LS concentrations were prepared initially and renewed daily. LS concentrations tested were 12.0, 17.0, 25.0, 35.0, and 50.0%. Test concentrations were determined from a range finding test that was initiated August 29, 2023 (Appendix G) This test was initiated September 23, 2013, at 14:23 and completed September 30, 2013, at 09:27.

MATERIALS AND METHODS

Materials and methods for the work performed are stated in EPA-821-R-02-014: Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms. Actual materials and methods are detailed below. This test was performed with strict adherence to the requirements of Method 1006 and/or the Western Gulf of Mexico OCS General Permit. The recommendations and suggestions made elsewhere in EPA-821-R-02-014 were incorporated whenever applicable to optimize the experimental design. Dilution water was prepared with hw-MARINEMIX + Bio-elements and Crystal Sea Marinemix Bioassay Laboratory Formula sea salts (80:20) and deionized water and adjusted to 25 parts per thousand (ppt) salinity.

M. beryllina was cultured and maintained at 24±1°C and 25 ppt salinity. Several clutches from different females comprised the embryo pool from which test organism population hatched. Test organisms were fed 200 – 300 µl of a standardized suspension of less than 24-hour-old *Artemia* nauplii twice daily by replicate. The standard suspension is equal to 0.05 g wet weight strained nauplii per ml synthetic seawater. Test organisms were not fed on Day 7. One day prior to test initiation, eight inland silverside minnows were transferred randomly into 30 test chambers with 250 ml synthetic seawater. These test chambers were then placed in the environmental chamber.

Sensitivity of test organisms to a known toxicant was determined by performing a chronic Standard Reference Toxicant (SRT) test, MN1310, with potassium chloride (Sigma Chemical, Lot SLBC2414V). The SRT test was initiated on September 04, 2013, with 11-day-old *M. beryllina* larvae. Appendix F contains *M. beryllina* SRT control charts.

	SURVIVAL	GROWTH
NOEC:	980 mg/L	980 mg/L
LOEC:	1400 mg/L	>980 mg/L

The sample used in this test was delivered to EE USA on September 20, 2013 (Appendix E). This sample was stored at 0.1 to 6°C and used to prepare the initial and subsequent renewal test solutions. Test chambers were labeled with replicate identification, and EE USA’s project number. Six treatments, five LS concentrations and a LPC were prepared and pH was measured in the undiluted LS sample daily (Appendix A, page 1).

Each treatment was poured into a new acid-washed 1-gallon plastic container and placed in an environmental chamber to warm up to test temperature. After the test solutions reached test temperature, initial water quality parameters (temperature, dissolved oxygen (DO), and salinity) were measured. At the end of each 24-hour exposure period, prior to renewal, the ending DO, temperature, salinity, and pH in each treatment were recorded also (Appendix A, pages 4 - 7). Alkalinity, pH, and salinity were measured in the LPC September 23, September 25, and September 27, 2013 (Appendix A, page 1).

On Day 0, the preloaded replicate test chambers were removed from the environmental chamber and carefully examined. Dead or injured larvae were replaced with organisms from the same batch and this test was initiated by renewal: excess food and debris was removed by pipette and 90% of the treatment solution was poured out of each replicate. Aliquots of freshly prepared treatments were poured gently into each replicate as appropriate and then this test was placed in the environmental chamber. Surviving test organisms were disturbed as little as possible during renewal. On Days 1 - 6, the test was renewed.

Every 24 hours, survival was recorded (Appendix A, pages 2 - 4). After seven days, the final survival data were recorded and this test was terminated. Surviving *M. beryllina* were rinsed in deionized water, placed on a tared weighing dish, and dried at 60+/-4°C for 24 hours by replicate. After cooling for at least 30 minutes, dried *M. beryllina* were weighed and the average individual dry weight for each replicate was calculated (Appendix C, page 3). The average individual dry weight is equal to the replicate weight divided by the number of original larvae.

Summary of Experimental Conditions

Test Organisms: 11-day-old *Menidia beryllina* larvae.
Dilution Water: Synthetic seawater, 25 ppt salinity.
Temperature: 25±1°C.
Photoperiod: 16 hours light; 8 hours dark.
Test Chambers: Rectangular Pyrex dish, 21 cm x 11 cm x 7 cm. Total volume = 1.45 L.
Test Solution Volume: 500 ml.
Aeration: Yes. On Day 1.
Test Solution Renewal: Yes.

Test acceptability criteria (TAC) include minimum LPC survival, 80%, minimum mean dry weight for surviving *M. beryllina* in the LPC, ≥0.50 mg, and maximum percent coefficient of variation (%CV) in the LPC and critical dilution for survival and growth, ≤40. The %CV was calculated using the number of surviving *M. beryllina* in each replicate. This test met all TAC. Survival in the concurrent LPC was 100.0%. The mean dry weight of surviving *M. beryllina* in the LPC was 1.148 mg and the highest %CV for survival and growth in the LPC and critical dilution was 6.4 (Appendix C, pages 2 & 4).

RESULTS AND CONCLUSION

The response used in statistical analysis of survival data was the proportion of surviving test organisms per replicate. These proportions were transformed by the Arc Sine Square Root Transformation and then tested for normal distribution and homogeneity of variance using Shapiro-Wilk's and Bartlett's tests, respectively. Survival data were normally distributed, unequal in variance, and further evaluated by the nonparametric alternative, Steel's Many-One Rank Test. The No Observed Effect Concentration (NOEC) for impaired *M. beryllina* survival was 25.0% LS. The Lowest Observed Effect Concentration (LOEC) was 35.0% LS. For this *M. beryllina* survival data set, the minimum statistically significant percent difference (MSDp) was 10.0 (Appendix C, page 2).

The response used in growth data analysis was the average individual dry weight for each replicate: replicate weight divided by the number of original larvae. Growth data were not transformed and concentrations demonstrating significant mortality are routinely excluded from subsequent data analysis. Growth data were tested for normal distribution and homogeneity of variance using Shapiro-Wilk's and Bartlett's tests, respectively. Growth data were normally distributed, equal in variance, and further evaluated by the parametric alternative, Dunnett's Test. The NOEC for impaired *M. beryllina* growth was <12.0% LS. The LOEC was 12.0% LS. For this *M. beryllina* growth data set, the MSDp was 15.5 (Appendix C, page 3).

Survival of *M. beryllina* larvae exposed to CHLOR*RID SP8 & CHLOR*RID SP8 RINSE was reduced significantly at 35.0% LS (the LOEC). Growth was reduced significantly at 12.0% LS (the LOEC). Survival and growth data summary statistics are presented in Appendix C.

MYSID (*Americamysis bahia*) SURVIVAL, GROWTH, AND FECUNDITY TEST
EPA-821-R-02-014: METHOD 1007

TEST OVERVIEW

A 7-day static-renewal toxicity test was conducted by EE USA to determine toxicity of LS CHLOR*RID SP8 & CHLOR*RID SP8 RINSE to *Americamysis bahia* juveniles. Methods, materials, and results are presented in this document. Organisms used in this test were cultured at EE USA and 7-days-old when this test was initiated. Synthetic seawater was used as the LPC solution and diluent in this test. Eight replicates of the LPC solution and five LS concentrations were prepared initially and renewed daily. LS concentrations tested were 0.6, 1.1, 1.8, 3.0, and 5.0%. Test concentrations tested were determined from a range finding test that was initiated August 29 and September 03, 2013 (Appendix G). This test was initiated September 23, 2013, at 14:22 and completed September 30, 2013, at 10:20.

MATERIALS AND METHODS

Materials and methods for the work performed are stated in EPA-821-R-02-014: Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms. Actual materials and methods are detailed below. This test was performed with strict adherence to the requirements of Method 1007 and/or the Western Gulf of Mexico OCS General Permit. The recommendations and suggestions made elsewhere in EPA-821-R-02-014 were incorporated whenever applicable to optimize the experimental design. Dilution water was prepared with hw-MARINEMIX + Bio-elements and Crystal Sea Marinemix Bioassay Laboratory Formula sea salts (80:20) and deionized water and adjusted to 25 ppt salinity.

A. bahia was cultured and maintained at 24±1°C and 25 ppt salinity. Six days before initiating this test, approximately 500, 12- to 24-hour-old mysids were collected from breeding cultures, moved to a holding system, and acclimated to 26±1°C. Test organisms were fed 100 – 175 µl of a standardized suspension of less than 24-hour-old *Artemia* nauplii twice daily by replicate. The standard suspension is equal to 0.05 g wet weight strained nauplii per ml synthetic seawater.

Sensitivity of test organisms to a known toxicant was determined by performing a chronic Standard Reference Toxicant (SRT) test AB1310, with potassium chloride (Sigma Chemical, Lot SLBC2414V). The SRT test was initiated on September 04, 2013, with 7-day-old *A. bahia*. Appendix F contains *A. bahia* SRT control charts.

	SURVIVAL	GROWTH
NOEC:	416 mg/L	416 mg/L
LOEC:	694 mg/L	>416 mg/L

The sample used in this test was delivered to EE USA on September 20, 2013 (Appendix E). This sample was stored at 0.1 to 6°C and used to prepare the initial and subsequent renewal test solutions. Test chambers were labeled with replicate identification, and EE USA's project number. Six treatments, five LS concentrations and a LPC were prepared and pH was measured in the undiluted LS sample daily (Appendix B, page 1).

Each treatment was poured into a new acid-washed 1-gallon plastic container and placed in an environmental chamber to warm up to test temperature. After the test solutions reached test temperature, initial water quality parameters (temperature, DO, and salinity) were measured. At the end of each 24-hour exposure period, prior to renewal, the ending DO, temperature, salinity, and pH in each treatment were recorded also (Appendix B, pages 4 - 7). Alkalinity, pH, and salinity were measured in the LPC September 23, September 25, and September 27, 2013 (Appendix B, page 1).

On Day 0, the treatments were poured into their respective test chambers, five *A. bahia* juveniles were distributed randomly to each, and then this test was placed in the environmental chamber. On Days 1 - 6, the test was renewed: excess food and debris was removed by pipette and 90% of the treatment solution was poured out of each replicate. Aliquots of freshly prepared treatments were poured gently into each replicate as appropriate. Surviving test organisms were disturbed as little as possible during renewal.

Every 24 hours, survival was recorded (Appendix B, pages 2 - 4). After seven days, the final survival data were recorded and this test was terminated. Surviving *A. bahia* were rinsed in deionized water, placed on a tared weighing dish, and dried at 60±4°C for 24 hours by replicate. After cooling for at least 30 minutes, dried *A. bahia* were weighed and the average individual dry weight for each replicate was calculated (Appendix D, page 4). The average individual dry weight is equal to the replicate weight divided by the number of original mysids.

Summary of Experimental Conditions

Test Organisms: 7-day-old *Americamysis bahia* juveniles.
Dilution Water: Synthetic seawater, 25 ppt salinity.
Temperature: 26±1°C.
Photoperiod: 16 hours light; 8 hours dark.
Test Chambers: Disposable plastic cups, 9 cm in diameter. Total volume = 300 ml.
Test Solution Volume: 150 ml.
Aeration: Yes. On Day 1.
Test Solution Renewal: Yes.

TAC include minimum LPC survival, 80%, minimum mean dry weight for surviving *A. bahia* in the LPC, ≥0.20 mg, and maximum %CV in the LPC and critical dilution for survival and growth, ≤40. The %CV was calculated using the number of surviving *A. bahia* in each replicate. This test met all TAC. Survival in the concurrent LPC was 100.0%. The mean dry weight of surviving *A. bahia* in the LPC was 0.322 mg and the highest %CV for survival and growth in the LPC and critical dilution was 14.9 (Appendix D, pages 3 & 5).

RESULTS AND CONCLUSION

The response used in statistical analysis of survival data was the proportion of surviving test organisms per replicate. These proportions were transformed by the Arc Sine Square Root Transformation and then tested for normal distribution and homogeneity of variance using Shapiro-Wilk's and Bartlett's tests, respectively. Survival data were not normally distributed and were further evaluated by the nonparametric alternative, Steel's Many-One Rank Test. The NOEC for impaired *A. bahia* survival was 1.1% LS. The LOEC was 1.8% LS. For this *A. bahia* survival data set, the MSDP was 13.4 (Appendix D, page 3).

The response used in growth data analysis was the average individual dry weight for each replicate: replicate weight divided by the number of original larvae. Growth data were not transformed and concentrations demonstrating significant mortality are routinely excluded from subsequent data analysis. Growth data were tested for normal distribution and homogeneity of variance using Shapiro-Wilk's and Bartlett's tests, respectively. Growth data were not normally distributed, equal in variance, and further evaluated by the nonparametric alternative, Steel's Many-One Rank Test. The NOEC for impaired *A. bahia* growth was 0.6% LS. The LOEC was 1.1% LS. For this *A. bahia* growth data set, the MSDp was 14.6 (Appendix D, page 4).

Survival of *A. bahia* exposed to CHLOR*RID SP8 & CHLOR*RID SP8 RINSE was reduced significantly at 1.8% LS (the LOEC). Growth was reduced significantly at 1.1% LS (the LOEC). Survival and growth data summary statistics are presented in Appendix D.

REFERENCES

- Environmental Enterprises USA, Quality Assurance Plan, June 2012 (or most recent version).
- Environmental Enterprises USA, Standard Operating Procedures, December 2012 (or most recent version).
- NELAC Institute. TNI Standard, Environmental Laboratory Sector, adopted September 8, 2009. Management and Technical Requirements for Laboratories Performing Environmental Analysis. Volume 1. EL-V1-2009-ISO. Weatherford, TX 76086.
- Tidepool Scientific Software. 2007. ToxCalc™ Toxicity Data Analysis Software. Version 5.0.32. McKinleyville, CA.
- U.S. Environmental Protection Agency. June 2000. Understanding and Accounting for Method Variability in Whole Effluent Toxicity Applications Under the National Pollutant Discharge Elimination System Program. EPA 833-R-00-003. Office of Wastewater Management (4203). Washington, DC 20460.
- U.S. Environmental Protection Agency. July 2000. Method Guidance and Recommendations for Whole Effluent Toxicity (WET) Testing (40 CFR Part 136). EPA 821-B-00-004. Office of Water (4303). Washington, DC 20460.
- U.S. Environmental Protection Agency. October 2002. Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms. EPA-821-R-02-014, Method 1006 and Method 1007. 3rd Edition. Office of Water (4303T). Washington, DC 20460.
- U.S. Environmental Protection Agency Region VI, Effective: October 1, 2012. Final NPDES General Permit for New and Existing Sources and New Dischargers in the Offshore Subcategory of the Oil and Gas Extraction Category for the Western Portion of the Outer Continental Shelf of the Gulf of Mexico (GMG290000). FR Volume 77, No. 196, p. 61605, October 10, 2012.

Environmental Enterprises USA, Inc.

APPENDIX A

CorrLine International – CorrX
Run-off from treated metal plate
 Barbara Tompkins-Brown

Test Concentrations, % Lab Sample (LS)

<i>Menidia beryllina</i>	Total Volume/ Concentration, ml	Color Code	ml LS	ml DH ₂ O
50.0	2500.00	Black	1250.00	1250.00
35.0	"	Red	875.00	1625.00
25.0	"	Yellow	625.00	1875.00
17.0	"	Green	425.00	2075.00
12.0	"	Blue	300.00	2200.00
0 LPC	"	White	0.00	2500.00
Total Volume (ml) of LS needed per day=				3475.00
Total Volume (ml) of LS needed for test duration=				24325.00

Data Pages & Calculations by: Veronica M... QA/QC Check by: Nick...

M. beryllina = 5 Reps x 500 ml
 = 2500 ml

DH₂O = Dilution Water = **Synthetic Seawater, 25 ppt**

	LPC	M #	LPC	M #	LPC	M #	LS #1	M #
Date	09/23		09/25		09/27		09/23	
Alkalinity	104	//	100	//	Ⓐ	//		//
Salinity	24.9	1B	25.3	1B	25.0	1B		
pH	8.0	A93	8.1	A93	8.0	A93		
	MUR		CMB		Ⓐ			

Artemia Lot #	
042012-2	
Initial	ME

LPC: Laboratory Performance Control, synthetic seawater
 Alkalinity: mg/l as CaCO₃ Salinity: ppt pH: su M#: meter number

Prep Date	09/23	09/24	09/25	09/26	09/27	09/28	09/29
DH ₂ O Lot #	25R-264-13	25R-265-13	25R-266-13	25R-267-13	25R-268-13	25R-269-13	25R-270-13
Sample #	1	1	1	1	1	1	1
pH	6.5	6.6	6.6	6.4	6.5	6.2	6.3
Meter #	A93	A93	A93	A93	A93	A93	A93
Initial	JA	CM	JA	CMB	JAG	JA	JA

Comments: Ⓐ Data Not Recorded 10/21/13 ME

Inland Silverside Minnow, *Menidia beryllina*
Larval Survival and Growth Test, Method 1006

CorrLine International – CorrX
Run-off from treated metal plate

Test Organisms Age: 11 Days Old Test Organisms Source: EE
 Test Initiation At: 1423 on 9/23/13
 Counted by: J. L. QC/QA by: Miranda Robbin
 Loaded by: Miranda Robbin Organism Lot #: MN-255-13
 Exposure Chamber: 1.5 L Pyrex dish. Feeding: *Artemia* nauplii 250 to 500 µl 2X / day / replicate.

***M. beryllina* Daily Survival Data**

Treatment: 0 % LS								White
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
1	8	8	8	8	8	8	8	8
2	8	8	8	8	8	8	8	8
3	8	8	8	8	8	8	8	8
4	8	8	8	8	8	8	8	8
5	8	8	8	8	8	8	8	8
Initials	MR	JA	CMB	Ag	SAG	#	#	SAG

Treatment: 12.0 % LS								Blue
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
6	8	8	7	7	7	7	7	7
7	8	8	7	7	7	7	7	7
8	8	8	8	8	8	8	7	7
9	8	8	8	7	7	7	7	7
10	8	8	8	8	8	8	8	8
Initials	MR	JA	CMB	Ag	SAG	#	#	SAG

Comments: _____

M. beryllina Daily Survival Data Cont.

Treatment: 17.0 % LS								Green
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
11	8	8	8	8	8	8	8	8
12	8	8	8	8	8	8	7	7
13	8	8	8	8	8	8	8	8
14	8	8	8	8	8	8	7	7
15	8	8	8	8	8	8	8	8
Initials	MR	JA	CMB	AJ	JAG	R	R	JAG

Treatment: 25.0 % LS								Yellow
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
16	8	8	8	8	8	7	7	7
17	8	8	7	7	7	6	5	5
18	8	8	8	8	8	7	6	6
19	8	8	8	8	8	8	8	8
20	8	8	8	7	6	5	5	5
Initials	MR	JA	CMB	AJ	JAG	R	R	JAG

Treatment: 35.0 % LS								Red
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
21	8	8	8	5	4	3	2	2
22	8	8	8	5	2	2	2	2
23	8	8	7	3	2	2	2	2
24	8	8	8	6	4	3	2	2
25	8	8	7	6	5	4	1	0
Initials	MR	JA	CMB	AJ	JAG	R	R	JAG

Comments: _____

M. beryllina Daily Survival Data Cont.

Treatment: 50.0 % LS								Black
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
26	8	8	8	0	0	0	0	0
27	8	8	6	0	0	0	0	0
28	8	8	6	0	0	0	0	0
29	8	8	5	0	0	0	0	0
30	8	8	4	2	0	0	0	0
Initials	mr	JA	CMB	Cy	JAG	pc	pc	JAG
Time	1423	1030	1121	1048	1040	0916	0955	0927

Test Completed on: 09/30/13

M. beryllina Water Quality Data

All Treatments: Initial Temp., 24.5 to 26.4°C. Final Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

Day 0	Treatment % LS							Comments
09/23/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO I	7.2	7.2	7.2	7.2	7.1	7.2	57	
Temp I	24.5	24.5	24.5	24.5	24.9	24.5	1B	
Salinity I	24.9	25.1	25.0	25.3	25.3	25.4	1B	
Tech Initials:		JA		Time:		1401		

Day 1	Treatment % LS							Comments
09/24/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO F	6.3	3.1	1.8	0.9	0.7	0.7	57	(A) wrong data 9/24/13, Cy
Temp F	24.9	24.8	24.9	25.0	24.8	24.9	1B	
Salinity F	25.3	25.3	25.3	25.3	25.8	25.5	1B	
pH F	8.0	7.6	7.5	7.5	7.5	7.4	AA3	
Tech Initials:		Cy		Time:		0837		

DO: mg/l pH: su Salinity: ppt Temp: °C

M. beryllina Water Quality Data Cont.

All Treatments: Initial Temp., 24.5 to 26.4°C. Final Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

Day 1		Treatment % LS						
09/24/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO	I	6.9	6.6	6.5	6.1	5.8	5.6	S7
Temp	I	25.2	24.9	24.6	24.9	24.8	25.0	IB
Salinity	I	25.1	25.2	25.2	25.3	25.3	25.4	IB
Tech Initials: CMB		Time: 0942						

Comments Aerated
at 39 mls per min

Day 2		Treatment % LS						
09/25/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO	F	7.1	6.6	6.0	5.1	4.7	5.4	S7
Temp	F	24.9	24.8	25.0	25.0	25.1	25.0	IB
Salinity	F	25.3	25.5	25.4	25.5	25.6	25.7	IB
pH	F	8.0	7.9	7.9	7.8	7.7	7.9	A93
Tech Initials: CMB		Time: 0839						

Comments _____

Day 2		Treatment % LS						
09/25/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO	I	7.3	6.9	6.8	6.2	6.0	5.2	S7
Temp	I	25.1	26.0	25.1	25.5	25.3	25.7	IB
Salinity	I	25.3	25.3	25.2	25.3	25.3	25.3	IB
Tech Initials: CMB		Time: 0933						

Comments _____

Day 3		Treatment % LS						
09/26/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO	F	6.8	6.3	4.8	5.9	5.2	5.3	S7
Temp	F	24.6	24.6	24.7	25.1	24.8	24.7	IB
Salinity	F	25.6	25.6	25.5	25.6	25.7	25.6	IB
pH	F	8.0	8.0	7.8	8.0	8.0	8.0	A93
Tech Initials: MFCM		Time: 0845						

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

M. beryllina Water Quality Data Cont.

All Treatments: Initial Temp., 24.5 to 26.4°C. Final Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

Day 3		Treatment % LS						
09/26/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO	I	7.1	6.9	6.8	6.5	6.1	5.5	
Temp	I	24.5	25.0	26.4	25.0	25.4	25.1	
Salinity	I	25.0	25.1	25.2	25.0	25.2	25.3	
Tech Initials:		CM CMB		Time: 1007				

Comments _____

Day 4		Treatment % LS						
09/27/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO	F	6.9	6.1	5.9	5.7	5.6	5.2	
Temp	F	25.2	25.0	25.0	25.0	25.1	25.0	
Salinity	F	25.1	25.5	25.5	25.6	25.6	25.7	
pH	F	8.0	8.1	8.1	8.2	8.1	8.0	
Tech Initials:		CM CMB		Time: 0732				

Comments _____

Day 4		Treatment % LS						
09/27/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO	I	7.2	7.0	7.0	6.9	6.5	6.0	
Temp	I	25.3	25.7	26.1	26.0	25.0	24.6	
Salinity	I	25.0	25.1	25.2	25.2	25.2	25.3	
Tech Initials:		CM CMB		Time: 0927				

Comments _____

Day 5		Treatment % LS						
09/28/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #	
DO	F	6.8	5.8	5.8	5.6	5.3	-	
Temp	F	24.8	25.1	24.6	24.6	24.7	-	
Salinity	F	25.3	25.3	25.5	25.5	25.5	-	
pH	F	8.0	8.1	8.2	8.1	8.1	-	
Tech Initials:		Ay		Time: 0739				

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

M. beryllina Water Quality Data Cont.

All Treatments: Initial Temp., 24.5 to 26.4°C. Final Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

Day 5		Treatment % LS					
09/28/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #
DO	I	7.0	6.9	6.9	6.6	6.5	57
Temp	I	25.1	24.5	24.5	25.1	25.5	1B
Salinity	I	25.0	24.9	25.0	25.0	25.0	1B
Tech Initials:		cy		Time:		0824	

Comments _____

Day 6		Treatment % LS					
09/29/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #
DO	F	6.8	6.2	5.2	5.2	5.4	57
Temp	F	24.8	24.5	24.7	24.8	24.7	1B
Salinity	F	25.4	25.4	25.4	25.3	25.5	1B
pH	F	8.0	8.0	8.0	8.2	8.2	A93
Tech Initials:		MECyg		Time:		0728	

Comments _____

Day 6		Treatment % LS					
09/29/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #
DO	I	7.1	6.9	6.7	6.4	6.4	57
Temp	I	24.8	24.8	25.1	24.6	24.8	1B
Salinity	I	24.9	25.0	25.0	25.0	25.1	1B
Tech Initials:		cy		Time:		0853	

Comments _____

Day 7		Treatment % LS					
09/30/13	LPC	12.0	17.0	25.0	35.0	50.0	Meter #
DO	F	7.0	5.8	6.2	6.1	5.7	57
Temp	F	24.4	24.6	24.7	24.7	24.6	1B
Salinity	F	25.3	25.4	25.3	25.4	25.6	1B
pH	F	8.1	8.2	8.2	8.2	8.2	A93
Tech Initials:		cy CMB		Time:		0828	

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

7 Day *M. beryllina* Growth Data

Rep #	Treatment % LS	A Final Weight (mg)	B Initial Weight (mg)	C No. of Orig. Larvae	D No. of Surv. Larvae
1	0	17.52	8.45	8	8
2	"	18.47	9.30	8	8
3	"	19.20	9.13	8	8
4	"	17.58	8.54	8	8
5	"	10.09	8.17	8	8
6	12.0	13.00	8.40	8	7
7	"	17.91	10.97	8	7
8	"	15.05	10.09	8	7
9	"	13.01	9.24	8	7
10	"	18.25	11.04	8	8
11	17.0	13.95	9.01	8	8
12	"	14.17	10.50	8	7
13	"	13.01	8.48	8	8
14	"	15.28	10.09	8	7
15	"	10.70	10.10	8	8

Comments: _____

7 Day *M. beryllina* Growth Data Cont.

Rep #	Treatment % LS	A Final Weight (mg)	B Initial Weight (mg)	C No. of Orig. Larvae	D No. of Surv. Larvae
16	25.0	11.70	8.09	8	7
17	"	12.01	9.70	8	5
18	"	11.31	9.50	8	6
19	"	13.90	9.70	8	8
20	"	13.52	10.08	8	5
21	35.0	8.87	7.15	8	2
22	"	9.70	8.97	8	2
23	"	10.08	9.09	8	2
24	"	8.72	8.00	8	2
25	"	—	9.21	8	0
26	50.0	—	8.18	8	0
27	"	—	9.01	8	0
28	"	—	9.00	8	0
29	"	—	9.08	8	0
30	"	—	7.95	8	0

Initial Foil Wts at 0834 on 9 / 27 / 2013 (wre) Scale#: R9

Oven Temp. 63.0 °C Therm. #: 7159

Begin Drying Survivors at 0927 on 09 / 30 / 2013 (JAG) Oven #: 14

Finish Drying Survivors at 0828 on 10 / 01 / 2013 (wre)

Final Foil Wts. at 0909 on 10 / 01 / 2013 (wre) Scale #: R9

Data Entry by: Vernica McNew

Double Data Entry by: Vernica McNew or

QA/QC Officer: Mara O'Hill

QA/QC Data Pages

- Company name & contact matches client file.
- Sample matches client file.
- Dilution series is correct:

12, 17, 25, 35, 50.

- Calculations on mixing page are correct. (sign mixing page)
- Dates, dilutions, test method, # of replicates, replicate volume, sample, acceptance limits, data analysis endpoint, and test organisms are correct throughout data pages.
- Format correct. (spaces for all entries, page numeration, no split pages, etc.)

ME Initials 9/23/13 Date

QA/QC Chain-of-Custody

ⓐ Section incomplete 100213 ME

- Sample on COC matches sample bottle.
- Sample on COC matches test data pages.
- Lab # on COC matches sample bottle.
- Lab # on COC matches test data pages.
- Sample volume is sufficient for test duration. (Sample volume in container(s) checked against sample volume on mixing page)

Sample volume available: _____ ml

Sample volume needed: _____ ml

(Sample volume insufficient if sample volume available < sample volume needed)

_____ Initials _____ Date

QA/QC Jugs & Labels

- Lab # on jug and labels matches test data pages.
- Dilution water type is on jug. (i.e. 25 ppt, 20 ppt, MHSF, etc.)
- Dilutions on jugs and labels match dilutions on test data pages.
- Jugs are color-coded. (see mixing page for appropriate color code sequence)

ME Initials 9/23/13 Date

QC/QA Raw Data: Microzein 10/2/13

Environmental Enterprises USA, Inc.

APPENDIX B

CorrLine International – CorrX
Run-off from treated metal plate
 Barbara Tompkins-Brown

Test Concentrations, % Produced Water (LS)

	<i>Mysidopsis bahia</i>	Total Volume/ Concentration, ml	Color Code	ml LS	ml DH ₂ O
	5.0	1200.00	Black	60.00	1140.00
	3.0	"	Red	36.00	1164.00
	1.8	"	Yellow	21.60	1178.40
	1.1	"	Green	13.20	1186.80
	0.6	"	Blue	7.20	1192.80
	0 LPC	"	White	0.00	1200.00
Total Volume (ml) of LS needed per day=					138.00
Total Volume (ml) of LS needed for test duration=					966.00

Data Pages & Calculations by: Veronica M. New QA/QC Check by: Michelle Zodin

M. bahia = 8 Reps x 150 ml
 = 1200 ml

DH₂O = Dilution Water = **Synthetic Seawater, 25 ppt**

	LPC	M #	LPC	M #	LPC	M #	LS #1	M #
Date	09/23		09/25		09/27		09/23	
Alkalinity	104	//	100	//	(B)	//		//
Salinity	24.9	1B	25.1	1B	25.0	1B		
pH	8.0	A93	8.1	A93	8.0	A93		
	MIR		CMB				(B)	

Artemia Lot #	
042012-2	
Initial	MW

LPC: Laboratory Performance Control, synthetic seawater
 Alkalinity: mg/l as CaCO₃ Salinity: ppt pH: su M#: meter number

Prep Date	09/23	09/24	09/25	09/26	09/27	09/28	09/29
DH ₂ O Lot #	25R-264-13	25R-265-13	25R-266-13	25R-267-13	25R-268-13	25R-269-13	25R-270-13
Sample #	1	1	1	1	1	1	1
pH	6.5	6.6	6.5	6.4	6.5	6.2	6.3
Meter #	A93	A93	A93	A93	A93	A93	A93
Initial	JA	CM	JA	@ ERAB JAG	JAG		

Comments: @Loring Data JAG 9-26-13
@Data not recorded 100213M7

Mysid, *Mysidopsis bahia*
Survival, Growth, and Fecundity Test, Method 1007

CorrLine International – CorrX
Run-off from treated metal plate

Test Organisms Age: 7 Days Old Test Organisms Source: EE
 Test Initiation At: 1422 on 9/23/13
 Counted by: Miranda Robbin QC/QA by: JR
 Loaded by: JR Organism Lot #: mb-508-13

Exposure Chamber: 300 ml plastic cup. **Feeding:** *Artemia* nauplii 150 to 250 µl 2X / day / replicate.

***M. bahia* Daily Survival Data**

Treatment: 0 % LS								White
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
1/2	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5
3/4	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5
5/6	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5
7/8	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5
Initials	JA	MR	JAG	CMB	JAG	JA	JA	MR

Treatment: 0.6 % LS								Blue
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
9/10	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5
11/12	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/4
13/14	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5
15/16	5/5	5/5	5/5	5/5	5/3	5/3	5/5	5/3
Initials	JA	MR	JAG	CMB	JAG	JA	JA	MR

Comments: _____

M. bahia Daily Survival Data Cont.

Treatment: 1.1 % LS								Green
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
17/18	5/5	5 15	5 15	5 15	5 15	5 15	5 15	4 15
19/20	5/5	5 15	5 15	5 15	5 15	5 15	5 15	4 15
21/22	5/5	5 15	5 15	5 15	5 15	5 15	5 15	5 15
23/24	5/5	5 15	5 15	5 15	5 15	5 15	5 15	5 15
Initials	JA	MR	JAG	CMB	JAG	JA	JA	MR

Treatment: 1.8 % LS								Yellow
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
25/26	5/5	5 15	5 15	5 15	5 15	5 15	5 15	2 14
27/28	5/5	5 15	5 15	5 15	5 15	5 15	3 15	1 13
29/30	5/5	5 15	5 15	5 15	5 15	5 15	5 15	2 10
31/32	5/5	5 15	5 15	5 15	5 15	5 15	5 15	4 14
Initials	JA	MR	JAG	CMB	JAG	JA	JA	MR

Treatment: 3.0 % LS							Ⓜ	Red
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
33/34	5/5	5 15	5 15	5 14	3 1 1	3 1 1	0 1 0	0 1 0
35/36	5/5	5 15	5 15	5 15	4 1 2	4 1 2	0 1 0	0 1 0
37/38	5/5	5 1 4	5 1 4	5 1 4	4 1 2	4 1 2	0 1 0	0 1 0
39/40	5/5	5 1 5	5 1 5	5 1 5	3 1 3	3 1 3	0 1 0	0 1 0
Initials	JA	MR	JAG	CMB	JAG	JA	JA	MR

Comments: Ⓜ wrong DATA 9-29-13 JA

M. bahia Daily Survival Data Cont.

Treatment: 5.0 % LS								Black
Rep.	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
41/42	5/5	5/5	5/5	0/2	0/0	0/0	0/0	0/0
43/44	5/5	5/5	5/5	3/3	0/0	0/0	0/0	0/0
45/46	5/5	5/5	5/5	2/2	0/0	0/0	0/0	0/0
47/48	5/5	5/5	5/5	1/4	0/0	0/0	0/0	0/0
Initials	JA	MR	SAG	CMR	SAG	0946	1013	MR
Time	1422	1039	1118	1156	1122	0946	11	1020

Test Completed on: 9/30/13

M. bahia Water Quality Data

All Treatments: Initial Temp., 24.5 to 26.4°C. Final Temp., 24.5 to 27.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

Day 0	Treatment % LS						
09/23/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO I	7.0	7.2	7.2	7.1	7.1	7.1	57
Temp I	24.5	24.8	24.6	24.8	25.0	24.8	1B
Salinity I	24.9	25.1	25.3	25.1	25.0	25.0	1B
Tech Initials: JA				Time: 1406			

Comments _____

Day 1	Treatment % LS						
09/24/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO F	5.8	5.6	5.0	4.6	3.2	2.0	57
Temp F	25.3	25.2	25.3	25.2	25.4	24.9	1B
Salinity F	26.1	25.8	26.0	26.0	26.0	26.0	1B
pH F	7.9	7.8	7.8	7.7	7.6	7.5	AA3
Tech Initials: Cy CMR				Time: 0833			

Comments TEST GRADUAL
0.39 mg/l/min on
Day 1 9/24/13 ure

DO: mg/l pH: su Salinity: ppt Temp: °C

M. bahia Water Quality Data Cont.

All Treatments: Initial Temp., 24.5 to 26.4°C. Final Temp., 24.5 to 27.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

Day 1		Treatment % LS							
09/24/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #	Comments _____	
DO	I	6.9	7.0	6.9	6.9	6.9	6.9	57	_____
Temp	I	25.4	25.5	25.5	25.4	25.4	25.4	1B	_____
Salinity	I	25.1	25.1	25.2	25.1	25.2	25.2	1B	_____
Tech Initials: CyCMB		Time: 0921							

Day 2		Treatment % LS							
09/25/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #	Comments _____	
DO	F	7.0	6.9	6.9	6.9	6.9	6.8	57	_____
Temp	F	25.0	24.8	25.1	25.1	25.3	24.8	1B	_____
Salinity	F	26.7	26.5	26.6	26.3	26.9	26.8	1B	_____
pH	F	8.1	8.1	8.1	8.1	8.2	8.2	A93	_____
Tech Initials: CyCMB		Time: 0841							

Day 2		Treatment % LS							
09/25/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #	Comments _____	
DO	I	7.2	7.2	7.1	7.1	7.1	7.1	57	_____
Temp	I	25.7	26.4	25.8	26.2	26.4	26.3	1B	_____
Salinity	I	25.1	25.1	25.1	25.0	25.1	25.1	1B	_____
Tech Initials: CMB CM		Time: 1003							

Day 3		Treatment % LS							
09/26/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #	Comments _____	
DO	F	6.7	6.8	6.8	6.9	6.8	6.8	57	_____
Temp	F	24.8	24.9	24.8	24.6	25.1	24.8	1B	_____
Salinity	F	26.7	26.5	26.6	26.5	26.7	26.5	1B	_____
pH	F	8.0	8.1	8.1	8.1	8.1	8.2	A93	_____
Tech Initials: MECM		Time: 0830							

DO: mg/l pH: su Salinity: ppt Temp: °C

M. bahia Water Quality Data Cont.

All Treatments: Initial Temp., 24.5 to 26.4°C. Final Temp., 24.5 to 27.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

Day 3		Treatment % LS						
09/26/13		LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO	I	7.1	7.0	7.1	7.1	7.1	7.0	S7
Temp	I	24.9	25.1	25.0	24.9	24.9	24.9	1B
Salinity	I	25.1	25.0	25.0	24.9	25.0	25.0	1B
Tech Initials:		CM CMB			Time:			0959

Comments _____

Day 4		Treatment % LS						
09/27/13		LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO	F	6.8	6.8	6.9	6.9	6.3	6.8	S7
Temp	F	25.2	25.1	25.0	25.0	24.8	24.8	1B
Salinity	F	26.8	26.4	26.9	26.4	27.1	26.5	1B
pH	F	8.1	8.1	8.1	8.1	7.9	8.2	ACB
Tech Initials:		CM CMB			Time:			0833

Comments _____

Day 4		Treatment % LS						
09/27/13		LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO	I	7.3	7.2	7.3	7.2	7.2	7.2	S7
Temp	I	24.7	24.7	24.7	24.8	24.7	24.9	1B
Salinity	I	25.0	25.1	25.1	25.0	25.0	25.1	1B
Tech Initials:		CM CMB			Time:			0920

Comments _____

Day 5		Treatment % LS						
09/28/13		LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO	F	6.8	6.7	6.8	6.6	6.7	6.7	S7
Temp	F	25.2	24.8	24.9	25.0	24.9	24.8	1B
Salinity	F	26.3	26.0	26.3	26.1	26.5	26.1	1B
pH	F	8.1	8.0	8.1	8.1	8.1	8.1	AA3
Tech Initials:		Cye			Time:			0727

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

M. bahia Water Quality Data Cont.

All Treatments: Initial Temp., 24.5 to 26.4°C. Final Temp., 24.5 to 27.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

Day 5	Treatment % LS						
09/28/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO I	7.0	6.9	6.9	6.9	6.9	-	57
Temp I	25.4	25.4	25.5	25.5	25.4	-	1B
Salinity I	25.0	25.0	25.0	25.0	25.0	-	1B
Tech Initials:		cy		Time:		0828	

Comments _____

Day 6	Treatment % LS						
09/29/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO F	7.0	7.0	7.0	6.9	-	-	57
Temp F	24.9	24.6	25.0	24.9	-	-	1B
Salinity F	27.0	27.1	26.7	26.7	-	-	1B
pH F	8.1	8.1	8.1	8.1	-	-	A93
Tech Initials:		ME cy		Time:		0913	

Comments _____

Day 6	Treatment % LS						
09/29/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO I	7.2	7.0	7.1	7.1	-	-	57
Temp I	24.6	24.7	24.6	24.8	-	-	1B
Salinity I	24.9	25.0	25.0	25.0	-	-	1B
Tech Initials:		cy		Time:		0824	

Comments _____

Day 7	Treatment % LS						
09/30/13	LPC	0.6	1.1	1.8	3.0	5.0	Meter #
DO F	6.7	6.7	6.8	6.8	-	-	57
Temp F	25.0	25.0	25.1	25.1	-	-	1B
Salinity F	27.1	26.8	27.1	26.7	-	-	1B
pH F	8.1	8.0	8.1	8.1	-	-	A93
Tech Initials:		cy CM15		Time:		0829	

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

CorrX, Run-off from treated metal plate

Q-626-13
 NOEC/LOEC

7 Day *M. bahia* Growth Data

Rep #	Treatment % LS	A Final Weight (mg)	B Initial Weight (mg)	C No. of Orig. Larvae	D No. of Surv. Larvae
1	0	11.04	9.45	5	5
2	"	10.13	9.05	5	5
3	"	10.64	8.80	5	5
4	"	8.85	7.30	5	5
5	"	10.51	8.07	5	5
6	"	10.02	8.94	5	5
7	"	9.47	7.80	5	5
8	"	9.10	7.49	5	5
9	0.6	8.90	7.03	5	5
10	"	8.52	6.41	5	5
11	"	8.31	6.00	5	5
12	"	7.31	5.63	5	4
13	"	9.74	7.92	5	5
14	"	9.43	7.80	5	5
15	"	9.14	7.35	5	5
16	"	7.94	6.84	5	3
17	1.1	9.30	7.95	5	4
18	"	9.09	7.57	5	5
19	"	8.51	7.18	5	4
20	"	7.14	6.06	5	5
21	"	8.33	6.93	5	5
22	"	8.91	7.03	5	5
23	"	9.33	7.82	5	5
24	"	9.07	7.00	5	5

7 Day *M. bahia* Growth Data Cont.

Rep #	Treatment % LS	A Final Weight (mg)	B Initial Weight (mg)	C No. of Orig. Larvae	D No. of Surv. Larvae
25	1.8	7.53	7.24	5	2
26	"	9.00	8.09	5	4
27	"	7.12	7.00	5	1
28	"	7.32	6.85	5	3
29	"	8.27	7.88	5	2
30	"	—	6.19	5	0
31	"	7.13	6.29	5	4
32	"	6.71	5.93	5	4
33	3.0	—	7.32	5	0
34	"	—	8.34	5	0
35	"	—	7.28	5	0
36	"	—	7.07	5	0
37	"	—	6.72	5	0
38	"	—	7.18	5	0
39	"	—	6.36	5	0
40	"	—	6.75	5	0
41	5.0	—	7.59	5	0
42	"	—	7.75	5	0
43	"	—	7.27	5	0
44	"	—	6.67	5	0
45	"	—	7.19	5	0
46	"	—	8.96	5	0
47	"	—	7.00	5	0
48	"	—	6.76	5	0

Initial Foil Wts at 0.440 on 9 / 27 / 2013 (MR) Scale#: R9

Oven Temp. 63.0 °C Therm. #: T159

Begin Drying Survivors at 1020 on 9 / 30 / 2013 (MR) Oven #: 1V

Finish Drying Survivors at 0.921 on 10 / 01 / 2013 (MR)

Final Foil Wts. at 1.343 on 10 / 01 / 2013 (MR) Scale #: R9

Data Entry by: Veronica McNew

Double Data Entry by: Veronica McNew or

QA/QC Officer: Maria O'Neil

Comments: _____

QA/QC Data Pages

- Company name & contact matches client file.
- Sample matches client file.
- Dilution series is correct:

0.6, 1.1, 1.8, 3.0, 5.0

- Calculations on mixing page are correct. (sign mixing page)
- Dates, dilutions, test method, # of replicates, replicate volume, sample, acceptance limits, data analysis endpoint, and test organisms are correct throughout data pages.
- Format correct. (spaces for all entries, page numeration, no split pages, etc.)

ME Initials 9/23/13 Date

QA/QC Chain-of-Custody

@section incomplete
100213 ME

- Sample on COC matches sample bottle.
- Sample on COC matches test data pages.
- Lab # on COC matches sample bottle.
- Lab # on COC matches test data pages.
- Sample volume is sufficient for test duration. (Sample volume in container(s) checked against sample volume on mixing page)

Sample volume available: _____ ml

Sample volume needed: _____ ml

(Sample volume insufficient if sample volume available < sample volume needed)

_____ Initials _____ Date

QA/QC Jugs & Labels

- Lab # on jug and labels matches test data pages.
- Dilution water type is on jug. (i.e. 25 ppt, 20 ppt, MHSF, etc.)
- Dilutions on jugs and labels match dilutions on test data pages.
- Jugs are color-coded. (see mixing page for appropriate color code sequence)

ME Initials 9/23/13 Date

QC/QA Raw Data:

Veronica Mc New

10/02/13

Environmental Enterprises USA, Inc.

APPENDIX C

Test: LF-Larval Fish Growth and Survival Test

Test ID: mn62513

Species: MB-Menidia beryllina

Protocol: EPAM 02-EPA Marine

Sample ID: GMG290000-NPDES Permit #

Sample Type: PRD-Product

Start Date: 9/23/2013 End Date: 9/30/2013

Lab ID: EE-Environmental Enterprises USA

Pos	ID	Rep	Group	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7	Total Wgt	Tare Wgt	Wgt Count
	1	1	LPC-LP Contro	8							8	17.52	8.45	8
	2	2	LPC-LP Contro	8							8	18.47	9.3	8
	3	3	LPC-LP Contro	8							8	19.26	9.13	8
	4	4	LPC-LP Contro	8							8	17.58	8.54	8
	5	5	LPC-LP Contro	8							8	16.69	8.17	8
	6	1	12	8							7	13.66	8.4	8
	7	2	12	8							7	17.91	10.97	8
	8	3	12	8							7	15.05	10.09	8
	9	4	12	8							7	13.61	9.24	8
	10	5	12	8							8	18.25	11.04	8
	11	1	17	8							8	13.95	9.01	8
	12	2	17	8							7	14.17	10.56	8
	13	3	17	8							8	13.61	8.48	8
	14	4	17	8							7	15.28	10.09	8
	15	5	17	8							8	16.7	10.1	8
	16	1	25	8							7	11.76	8.09	8
	17	2	25	8							5	12.01	9.7	8
	18	3	25	8							6	11.31	9.5	8
	19	4	25	8							8	13.96	9.7	8
	20	5	25	8							5	13.52	10.08	8
	21	1	35	8							2	8.87	7.15	8
	22	2	35	8							2	9.7	8.97	8
	23	3	35	8							2	10.68	9.69	8
	24	4	35	8							2	8.72	8.06	8
	25	5	35	8							0			8
	26	1	50	8							0			8
	27	2	50	8							0			8
	28	3	50	8							0			8
	29	4	50	8							0			8
	30	5	50	8							0			8

Comments:

Larval Fish Growth and Survival Test-7 Day Survival

Start Date: 9/23/2013	Test ID: mn62513	Sample ID: GMG290000-NPDES Permit #
End Date: 9/30/2013	Lab ID: EE-Environmental Enterprise	Sample Type: PRD-Product
Sample Date:	Protocol: EPAM 02-EPA Marine	Test Species: MB-Menidia beryllina
Comments:		

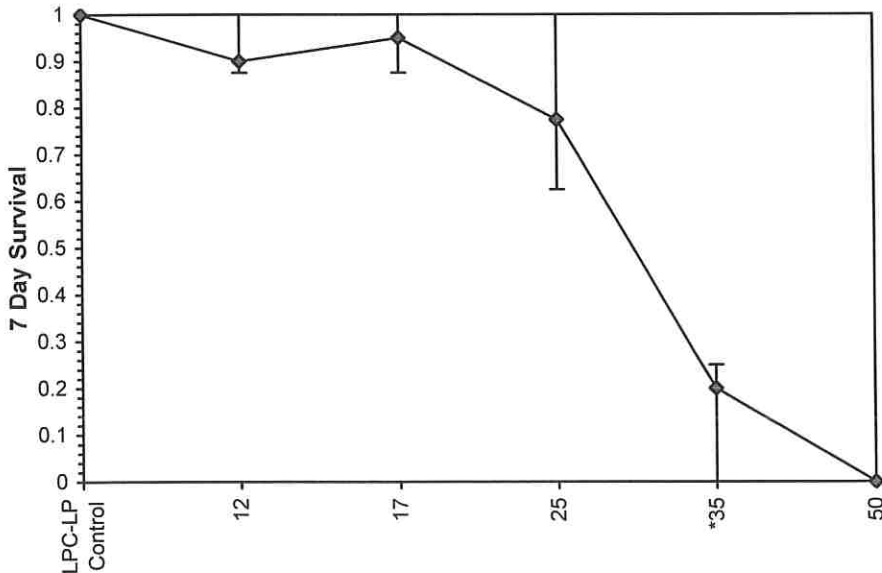
Conc-%	1	2	3	4	5
PC-LP Control	1.0000	1.0000	1.0000	1.0000	1.0000
12	0.8750	0.8750	0.8750	0.8750	1.0000
17	1.0000	0.8750	1.0000	0.8750	1.0000
25	0.8750	0.6250	0.7500	1.0000	0.6250
35	0.2500	0.2500	0.2500	0.2500	0.0000
50	0.0000	0.0000	0.0000	0.0000	0.0000

Conc-%	Mean	N-Mean	Transform: Arcsin Square Root				N	Rank Sum	1-Tailed Critical
			Mean	Min	Max	CV%			
PC-LP Control	1.0000	1.0000	1.3931	1.3931	1.3931	0.000	5		
12	0.9000	0.9000	1.2462	1.2094	1.3931	6.591	5	17.50	17.00
17	0.9500	0.9500	1.3196	1.2094	1.3931	7.623	5	22.50	17.00
25	0.7750	0.7750	1.0946	0.9117	1.3931	18.911	5	17.50	17.00
*35	0.2000	0.2000	0.4544	0.1777	0.5236	34.040	5	15.00	17.00
50	0.0000	0.0000	0.1777	0.1777	0.1777	0.000	5		

Auxiliary Tests	Statistic	Critical	Skew	Kurt
Shapiro-Wilk's Test indicates normal distribution (p > 0.05)	0.95154	0.918	-0.0722	1.31124
Equality of variance cannot be confirmed				

Hypothesis Test (1-tail, 0.05)	NOEC	LOEC	ChV	TU
Steel's Many-One Rank Test	25	35	29.5804	4
Treatments vs LPC-LP Control				

Dose-Response Plot



Hypothesis Test (1-tail, 0.05)	NOEC	LOEC	ChV	TU	MSDu	MSDp	MSB	MSE	F-Prob	df
Dunnett's Test	17	25	20.6155	5.88235	0.09673	0.09985	0.71536	0.01673	1.6E-09	4, 20
Treatments vs LPC-LP Control										

Larval Fish Growth and Survival Test-7 Day Growth

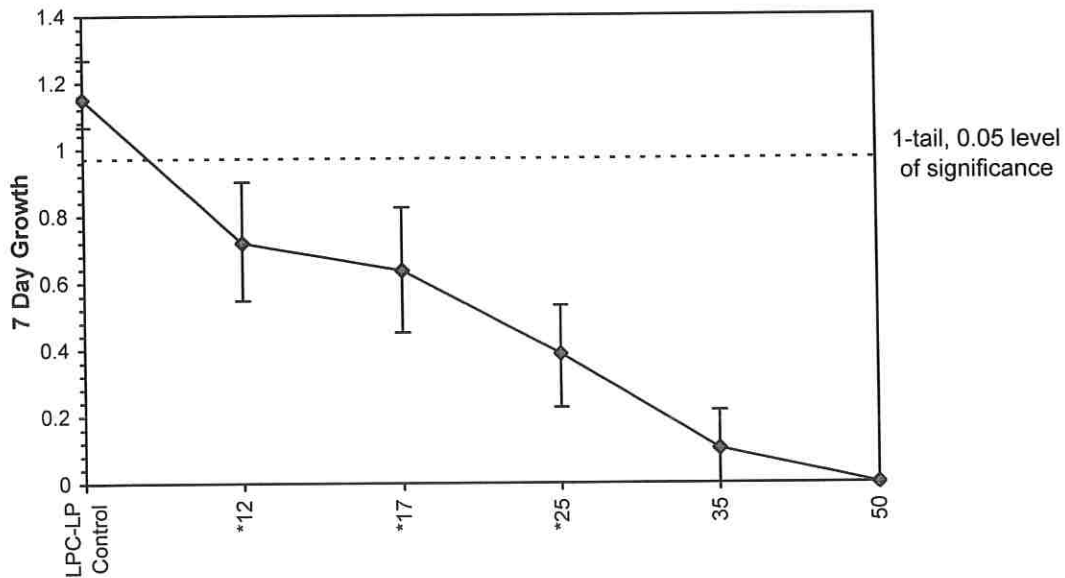
Start Date: 9/23/2013	Test ID: mn62513	Sample ID: GMG290000-NPDES Permit #
End Date: 9/30/2013	Lab ID: EE-Environmental Enterprise	Sample Type: PRD-Product
Sample Date:	Protocol: EPAM 02-EPA Marine	Test Species: MB-Menidia beryllina
Comments:		

Conc-%	1	2	3	4	5
PC-LP Control	1.1338	1.1463	1.2663	1.1300	1.0650
12	0.6575	0.8675	0.6200	0.5463	0.9013
17	0.6175	0.4513	0.6413	0.6488	0.8250
25	0.4588	0.2888	0.2263	0.5325	0.4300
35	0.2150	0.0912	0.1238	0.0825	0.0000
50	0.0000	0.0000	0.0000	0.0000	0.0000

Conc-%	Mean	N-Mean	Transform: Untransformed					N	t-Stat	1-Tailed	
			Mean	Min	Max	CV%	Critical			MSD	
PC-LP Control	1.1483	1.0000	1.1483	1.0650	1.2663	6.370	5				
*12	0.7185	0.6257	0.7185	0.5463	0.9013	21.862	5	5.392	2.230	0.1777	
*17	0.6368	0.5545	0.6368	0.4513	0.8250	20.832	5	6.417	2.230	0.1777	
*25	0.3873	0.3373	0.3873	0.2263	0.5325	32.577	5	9.547	2.230	0.1777	
35	0.1025	0.0893	0.1025	0.0000	0.2150	75.791	5				
50	0.0000	0.0000	0.0000	0.0000	0.0000	0.000	5				

Auxiliary Tests	Statistic	Critical	Skew	Kurt						
Shapiro-Wilk's Test indicates normal distribution (p > 0.05)	0.95355	0.905	0.10724	-0.9068						
Bartlett's Test indicates equal variances (p = 0.58)	1.96827	11.3449								
Hypothesis Test (1-tail, 0.05)	NOEC	LOEC	ChV	TU	MSDu	MSDp	MSB	MSE	F-Prob	df
Dunnett's Test	<12	12			0.17775	0.1548	0.50171	0.01588	5.9E-07	3, 16
Treatments vs LPC-LP Control										

Dose-Response Plot



Larval Fish Growth and Survival Test-7 Day Growth

Start Date: 9/23/2013	Test ID: mn62513cv	Sample ID: GMG290000-NPDES Permit #
End Date: 9/30/2013	Lab ID: EE-Environmental Enterprise	Sample Type: PRD-Product
Sample Date:	Protocol: EPAM 02-EPA Marine	Test Species: MB-Menidia beryllina
Comments:		

Conc-%	1	2	3	4	5
PC-LP Control	1.1338	1.1463	1.2663	1.1300	1.0650
12	0.7514	0.9914	0.7086	0.6243	0.9013
17	0.6175	0.5157	0.6413	0.7414	0.8250
25	0.5243	0.4620	0.3017	0.5325	0.6880
35	0.8600	0.3650	0.4950	0.3300	

Conc-%	Mean	N-Mean	Transform: Untransformed				N
			Mean	Min	Max	CV%	
PC-LP Control	1.1483	1.0000	1.1483	1.0650	1.2663	6.370	5
12	0.7954	0.6927	0.7954	0.6243	0.9914	18.689	5
17	0.6682	0.5819	0.6682	0.5157	0.8250	17.787	5
25	0.5017	0.4369	0.5017	0.3017	0.6880	27.791	5
35	0.5125	0.4463	0.5125	0.3300	0.8600	47.278	4

Auxiliary Tests	Statistic	Critical	Skew	Kurt
Shapiro-Wilk's Test indicates normal distribution (p > 0.05)	0.95407	0.916	0.68572	0.44254
Bartlett's Test indicates equal variances (p = 0.33)	4.57127	13.2767		

Environmental Enterprises USA, Inc.

APPENDIX D

Test: ACU-Acute Toxicity Test
 Species: MY-Mysidopsis bahia
 Sample ID: GMG290000-NPDES Permit #
 Start Date: 9/23/2013 End Date: 9/30/2013
 Test ID: mb62613
 Protocol: EPA 02-EPA Acute
 Sample Type: PRD-Product
 Lab ID: EE-Environmental Enterprises USA

Pos ID	Rep	Group	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7	Total Wgt((Tare Wgt((Wgt Count	Female Cc	Females w/ Eggs	Notes
1	1	LPC-LP Contro	5							5	11.04	9.45	5	
2	2	LPC-LP Contro	5							5	10.13	9.05	5	
3	3	LPC-LP Contro	5							5	10.64	8.8	5	
4	4	LPC-LP Contro	5							5	8.85	7.3	5	
5	5	LPC-LP Contro	5							5	10.51	8.67	5	
6	6	LPC-LP Contro	5							5	10.62	8.94	5	
7	7	LPC-LP Contro	5							5	9.47	7.8	5	
8	8	LPC-LP Contro	5							5	9.1	7.49	5	
9	1	0.6	5							5	8.9	7.03	5	
10	2	0.6	5							5	8.52	6.41	5	
11	3	0.6	5							5	8.31	6.6	5	
12	4	0.6	5							4	7.31	5.63	5	
13	5	0.6	5							5	9.74	7.92	5	
14	6	0.6	5							5	9.43	7.8	5	
15	7	0.6	5							5	9.14	7.35	5	
16	8	0.6	5							3	7.94	6.84	5	
17	1	1.1	5							4	9.3	7.95	5	
18	2	1.1	5							5	9.09	7.57	5	
19	3	1.1	5							4	8.51	7.18	5	
20	4	1.1	5							5	7.14	6.06	5	
21	5	1.1	5							5	8.33	6.93	5	
22	6	1.1	5							5	8.91	7.63	5	
23	7	1.1	5							5	9.33	7.82	5	
24	8	1.1	5							5	9.07	7.6	5	
25	1	1.8	5							2	7.53	7.24	5	
26	2	1.8	5							4	9	8.09	5	
27	3	1.8	5							1	7.12	7	5	
28	4	1.8	5							3	7.32	6.85	5	
29	5	1.8	5							2	8.27	7.88	5	
30	6	1.8	5							0			5	
31	7	1.8	5							4	7.13	6.29	5	
32	8	1.8	5							4	6.71	5.93	5	
33	1	3	5							0			5	
34	2	3	5							0			5	
35	3	3	5							0			5	
36	4	3	5							0			5	
37	5	3	5							0			5	
38	6	3	5							0			5	

Test: ACU-Acute Toxicity Test Species: MY-Mysidopsis bahia Sample ID: GMG290000-NPDES Permit # Start Date: 9/23/2013 End Date: 9/30/2013 Test ID: mb62613 Protocol: EPAA 02-EPA Acute Sample Type: PRD-Product Lab ID: EE-Environmental Enterprises USA															
Pos ID	Rep	Group	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7	Total Wgt(Tare Wgt)	(Wgt Count)	Female Cc	Females w/ Eggs	Notes
39	7		3	5						0		5			
40	8		3	5						0		5			
41	1		5	5						0		5			
42	2		5	5						0		5			
43	3		5	5						0		5			
44	4		5	5						0		5			
45	5		5	5						0		5			
46	6		5	5						0		5			
47	7		5	5						0		5			
48	8		5	5						0		5			

Comments:

Acute Toxicity Test-7 Day Survival

Start Date: 9/23/2013	Test ID: mb62613	Sample ID: GMG290000-NPDES Permit #
End Date: 9/30/2013	Lab ID: EE-Environmental Enterprise	Sample Type: PRD-Product
Sample Date:	Protocol: EPAA 02-EPA Acute	Test Species: MY-Mysidopsis bahia
Comments:		

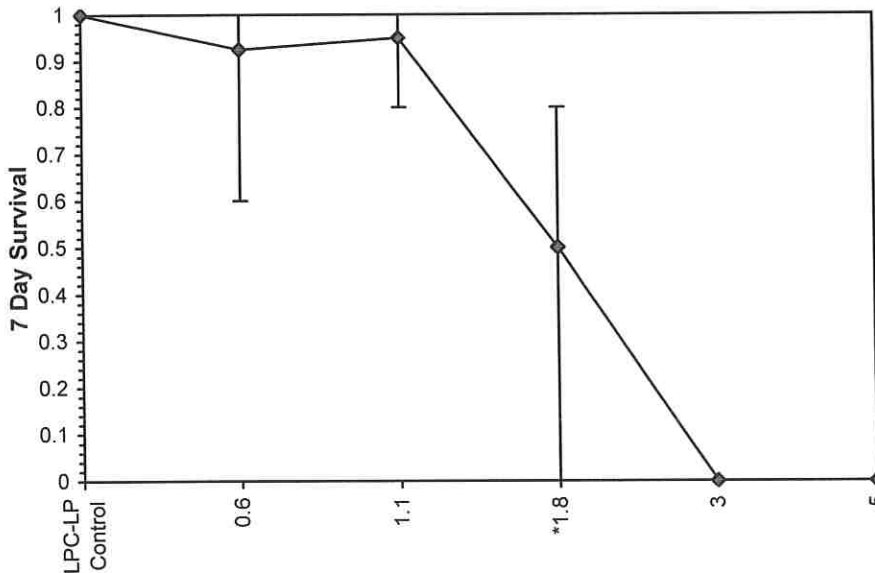
Conc-%	1	2	3	4	5	6	7	8
PC-LP Control	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000
0.6	1.0000	1.0000	1.0000	0.8000	1.0000	1.0000	1.0000	0.6000
1.1	0.8000	1.0000	0.8000	1.0000	1.0000	1.0000	1.0000	1.0000
1.8	0.4000	0.8000	0.2000	0.6000	0.4000	0.0000	0.8000	0.8000
3	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000
5	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000

Conc-%	Mean	N-Mean	Transform: Arcsin Square Root					Rank Sum	1-Tailed Critical
			Mean	Min	Max	CV%	N		
PC-LP Control	1.0000	1.0000	1.3453	1.3453	1.3453	0.000	8		
0.6	0.9250	0.9250	1.2581	0.8861	1.3453	13.661	8	60.00	48.00
1.1	0.9500	0.9500	1.2857	1.1071	1.3453	8.574	8	60.00	48.00
*1.8	0.5000	0.5000	0.7833	0.2255	1.1071	42.011	8	36.00	48.00
3	0.0000	0.0000	0.2255	0.2255	0.2255	0.000	8		
5	0.0000	0.0000	0.2255	0.2255	0.2255	0.000	8		

Auxiliary Tests	Statistic	Critical	Skew	Kurt
Shapiro-Wilk's Test indicates non-normal distribution (p <= 0.05)	0.88118	0.93	-0.9174	2.23982
Equality of variance cannot be confirmed				

Hypothesis Test (1-tail, 0.05)	NOEC	LOEC	ChV	TU
Steel's Many-One Rank Test	1.1	1.8	1.40712	90.9091
Treatments vs LPC-LP Control				

Dose-Response Plot



Hypothesis Test (1-tail, 0.05)	NOEC	LOEC	ChV	TU	MSDu	MSDp	MSB	MSE	F-Prob	df
Dunnett's Test	1.1	1.8	1.40712	90.9091	0.12699	0.13367	0.53716	0.03749	7.6E-06	3, 28
Treatments vs LPC-LP Control										

Acute Toxicity Test-7 Day Growth

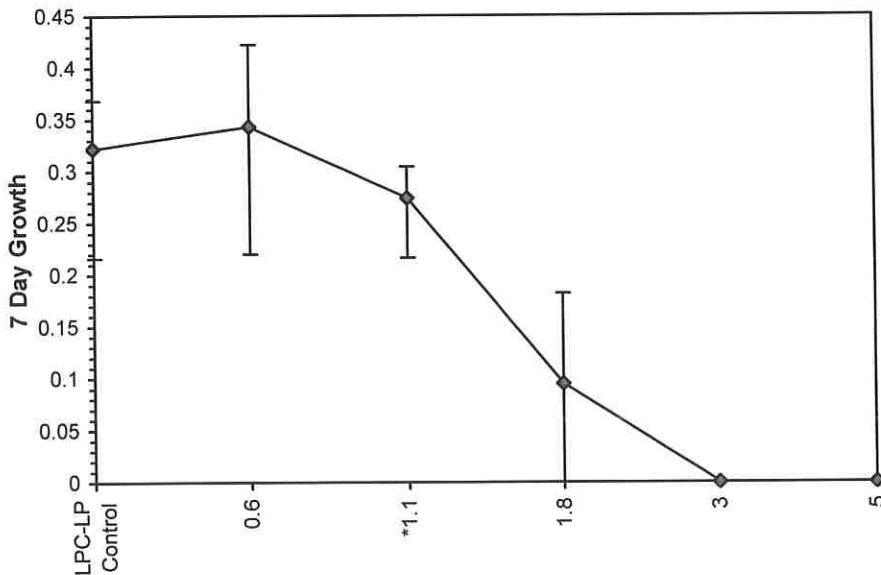
Start Date: 9/23/2013	Test ID: mb62613	Sample ID: GMG290000-NPDES Permit #
End Date: 9/30/2013	Lab ID: EE-Environmental Enterprise	Sample Type: PRD-Product
Sample Date:	Protocol: EPAA 02-EPA Acute	Test Species: MY-Mysidopsis bahia
Comments:		

Conc-%	1	2	3	4	5	6	7	8
PC-LP Control	0.3180	0.2160	0.3680	0.3100	0.3680	0.3360	0.3340	0.3220
0.6	0.3740	0.4220	0.3420	0.3360	0.3640	0.3260	0.3580	0.2200
1.1	0.2700	0.3040	0.2660	0.2160	0.2800	0.2560	0.3020	0.2940
1.8	0.0580	0.1820	0.0240	0.0940	0.0780	0.0000	0.1680	0.1560
3	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000
5	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000

Conc-%	Mean	N-Mean	Transform: Untransformed					Rank Sum	1-Tailed Critical
			Mean	Min	Max	CV%	N		
PC-LP Control	0.3215	1.0000	0.3215	0.2160	0.3680	14.855	8		
0.6	0.3428	1.0661	0.3428	0.2200	0.4220	16.845	8	80.50	49.00
*1.1	0.2735	0.8507	0.2735	0.2160	0.3040	10.602	8	43.50	49.00
1.8	0.0950	0.2955	0.0950	0.0000	0.1820	71.552	8		
3	0.0000	0.0000	0.0000	0.0000	0.0000	0.000	8		
5	0.0000	0.0000	0.0000	0.0000	0.0000	0.000	8		

Auxiliary Tests	Statistic	Critical	Skew	Kurt
Shapiro-Wilk's Test indicates non-normal distribution (p <= 0.05)	0.87321	0.916	-1.3172	2.61852
Bartlett's Test indicates equal variances (p = 0.23)	2.91989	9.21034		
Hypothesis Test (1-tail, 0.05)	NOEC	LOEC	ChV	TU
Steel's Many-One Rank Test	0.6	1.1	0.8124	166.667
Treatments vs LPC-LP Control				

Dose-Response Plot



Hypothesis Test (1-tail, 0.05)	NOEC	LOEC	ChV	TU	MSDu	MSDp	MSB	MSE	F-Prob	df
Dunnett's Test	0.6	1.1	0.8124	166.667	0.04695	0.14604	0.01007	0.00215	0.02087	2, 21
Treatments vs LPC-LP Control										

Acute Toxicity Test-7 Day Growth

Start Date: 9/23/2013	Test ID: mb62613cv	Sample ID: GMG290000-NPDES Permit #
End Date: 9/30/2013	Lab ID: EE-Environmental Enterprise	Sample Type: PRD-Product
Sample Date:	Protocol: EPAA 02-EPA Acute	Test Species: MY-Mysidopsis bahia

Conc-%	1	2	3	4	5	6	7	8
PC-LP Control	0.3180	0.2160	0.3680	0.3100	0.3680	0.3360	0.3340	0.3220
0.6	0.3740	0.4220	0.3420	0.4200	0.3640	0.3260	0.3580	0.3667
1.1	0.3375	0.3040	0.3325	0.2160	0.2800	0.2560	0.3020	0.2940
1.8	0.1450	0.2275	0.1200	0.1567	0.1950	0.2100	0.1950	

Transform: Untransformed

Conc-%	Mean	N-Mean	Mean	Min	Max	CV%	N
PC-LP Control	0.3215	1.0000	0.3215	0.2160	0.3680	14.855	8
0.6	0.3716	1.1558	0.3716	0.3260	0.4220	9.161	8
1.1	0.2903	0.9028	0.2903	0.2160	0.3375	13.750	8
1.8	0.1785	0.5551	0.1785	0.1200	0.2275	21.654	7

Auxiliary Tests	Statistic	Critical	Skew	Kurt
Shapiro-Wilk's Test indicates normal distribution (p > 0.05)	0.93636	0.929	-0.7603	0.62603
Bartlett's Test indicates equal variances (p = 0.85)	0.78367	11.3449		

Environmental Enterprises USA, Inc.

APPENDIX E

ENVIRONMENTAL ENTERPRISES USA, INC.

58485 Pearl Acres Rd., Suite D
Slidell, Louisiana 70461

(985) 646-2787

Copy Initiated by T. O'Reilly
EE on 9-20-13 @ 1040

158 A TO
Kit No. 157 A

CHAIN - OF - CUSTODY RECORD

Client: Corr Line Contact Person: Barbara Tompkins-Brown Special Handling _____
 Address: _____ Phone#: _____ Request _____
 P.O. # _____ () RUSH _____
 FAX #: _____ () VERBAL _____
 Project: _____ () OTHER _____

Lab Sample Description	Date Collected	Time Collected	No. of Containers	Analysis Request	SIR No.	Lab No.
Corr X			4	Mu/Mb chronic		0-625-13 m 0-626-13 mb

Collected By:	Date	Time	Relinquished By:	Date	Time
Received By: <u>T. O'Reilly</u>	9-20-13	1040	Relinquished By: <u>T. O'Reilly EE</u>	9-20-13	1045
Received By: <u>Veronica McNew</u>	09/20/13	1047	Relinquished By:		
Received By:	Date	Time	Relinquished By:	Date	Time
Received By:	Date	Time	Relinquished By:	Date	Time

J. EA 8995 9499 7491
 8995 8499 7480

SEARCHED
 10/14/13 mm

Kit # 158A 70

FedEx NEW Package Express US Airbill
Tracking Number 8995 9499 7491

1 From this portion can be removed for recipient's records.
 Date 9/19/85 FedEx Tracking Number 899594997491
 Sender's Name Kink Chrisman Phone
 Company Caroline Int'l
 Address 8 Glenway Blvd. # 910
 City Houston TX ZIP 77046

2 Your Internal Billing Reference
 3 To Recipient's Name SHIPPING AND RECEIVING Phone 955 645-2787

Company ENVIRONMENTAL ENTERPRISES USA
 Address 58485 PEARL ACRES RD STE 2
 We cannot deliver to P.O. boxes or P.O. ZIP codes.
 Address SLIDELL LA ZIP 70448-5400
 Use this line for the HOLD location address or for continuation of your shipping address.
 City SLIDELL State LA ZIP 70448-5400

0100055469
 8995 9499 7491



4 Express Package Service *To most locations. For packages over 150 lbs, use the new FedEx Express Freight US Airbill.
 NOTE: Service order has changed! Please select carefully.
 Next Business Day
 FedEx First Overnight
 FedEx Priority Overnight
 FedEx Standard Overnight
 FedEx 2Day
 FedEx 2Day AM
 FedEx Express Saver

5 Packaging *Declared value limit \$500.
 FedEx Envelope* FedEx Pak* FedEx Box FedEx Tube Other

6 Special Handling and Delivery Signature Options
 SATURDAY Delivery
 NOT available for FedEx Standard Overnight, FedEx 2Day A.M., or FedEx Express Saver.

No Signature Required
 Direct Signature
 Indirect Signature
 Does this shipment contain dangerous goods?
 No Yes Restricted
 Yes Shipper's Declaration
 Yes Shipper's Declaration
 Dry Ice Dry Ice
 Cargo Aircraft Only

7 Payment Bill to
 Enter FedEx Acct. No. or Credit Card No. below.
 Sender Recipient Third Party
 Acct. No. Shown Total Bill
 Obtain recip. Acct. No. Credit Card Cash/Check
 Total Packages Total Weight
 Credit Card Auth.

13111
1500

fedex.com 1800.GoFedEx 1800.463.3339

Rev. Date 1/10 - Per #10314 - ©1985-2010 FedEx - PRINTED IN U.S.A. SBS

Kit # 157A

1314

FedEx NEW Package Express **US Airbill** Tracking Number 8995 9499 7480

1 From Tracking Number can be removed for recipient's records. **91913** FedEx Tracking Number **8995 9499 7480**

Date **9/19/13**

Sender's Name **Kirk Chrisman** Phone _____

Company **Carline Int'l**

Address **8 Glenway Blvd. #910**

City **Houston** State **TX** ZIP **77046** Dist./P.O./Sub./Room _____

2. Your Internal Billing Reference

3 To Recipient's SHIPPING AND RECEIVING Name **ENVIRONMENTAL ENTERPRISES USA** Phone **985 646-2787**

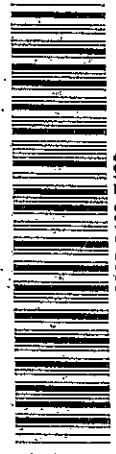
Address **5885 PEARL ACRES RD STE D** Dist./P.O./Sub./Room _____

We cannot deliver to P.O. boxes or P.O. ZIP codes.

Address _____

Use this line for the HOLD location address or for continuation of your shipping address.

City **SLIDELL** State **LA** ZIP **70461-5400**



8995 9499 7480



4 Express Package Service. Packages up to 150 lbs. For packages over 20 lbs., use the new FedEx Express Freight & Adult.

Next Business Day Earliest next business morning delivery to select business locations. Packages will be delivered on Monday, Tuesday, Wednesday, Thursday or Friday, subject to availability.

FedEx First Overnight Earliest next business morning delivery to select business locations. Packages will be delivered on Monday, Tuesday, Wednesday, Thursday or Friday, subject to availability.

FedEx Priority Overnight Next business morning. Friday shipments will be delivered on Monday unless SATURDAY Delivery is selected.

FedEx Standard Overnight Next business afternoon. Saturday Delivery NOT available.

2 or 3 Business Days NEW FedEx 2Day A.M. Second business morning. Saturday Delivery NOT available.

FedEx 2Day Second business afternoon. Thursday shipments will be delivered on Monday unless SATURDAY Delivery is selected.

FedEx Express Saver Third business day. Saturday Delivery NOT available.

5 Packaging *Declared value limit \$2k.

FedEx Envelope FedEx Pak FedEx Box FedEx Tube Other

6 Special Handling and Delivery Signature Options

SATURDAY Delivery NOT available for FedEx Standard Overnight, FedEx 2Day A.M., or FedEx Express Saver.

No Signature Required Package may be left without obtaining a signature for delivery.

Direct Signature Someone at recipient's address may sign for delivery. Fee applies.

Indirect Signature (from one to another) address, someone at a neighbor's address may sign for delivery. Recipiental delivery only. Applies.

Does this shipment contain dangerous goods? No. Yes. This box must be checked.

Yes. Hazardous materials. Shipper's Declaration and label required.

Yes. Perishable. Shipper's Declaration and label required.

Dry Ice Dry Ice 3, UN 1845 Cargo Aircraft Only

7 Payment Bill to

Sender Recipient Third Party Credit Card Cash/Check

Account to be billed _____

Enter FedEx Acct. No. or Credit Card No. below _____

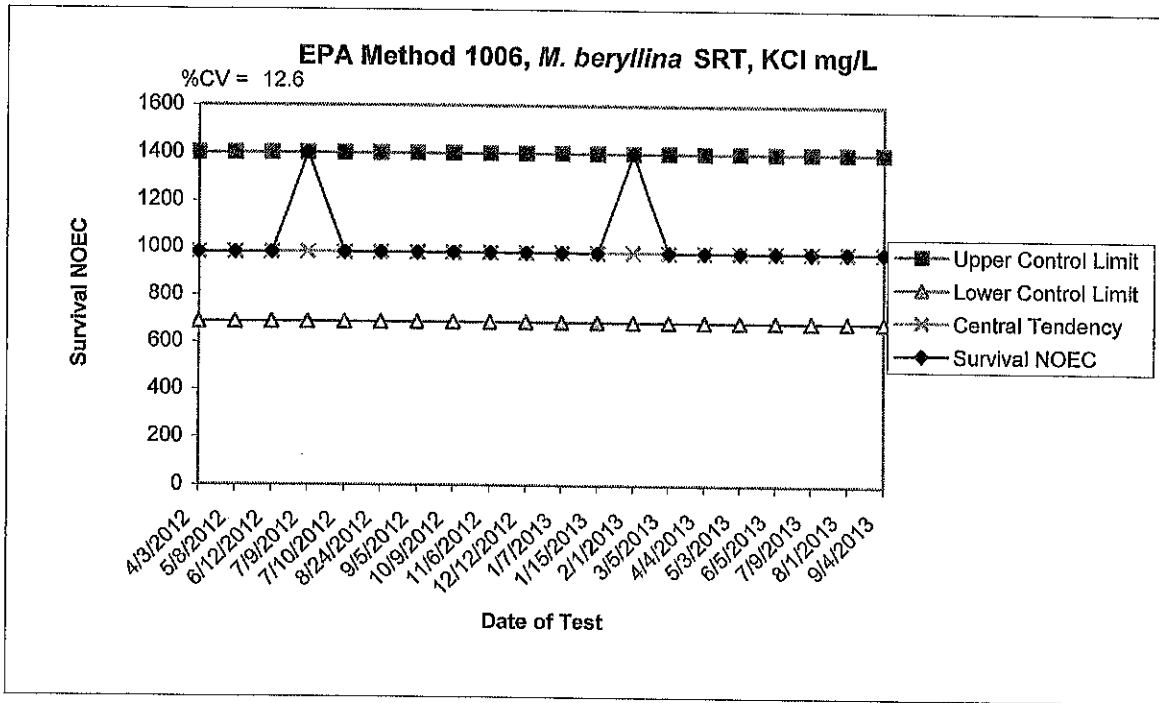
Total Packages **1** Total Weight **34** lbs.

Credit Card Auth. **6111**

Rev. Date 11/10 - Parcels 13 - ©1995-2010 FedEx - PRINTED IN U.S.A. SIS

Environmental Enterprises USA, Inc.

APPENDIX F



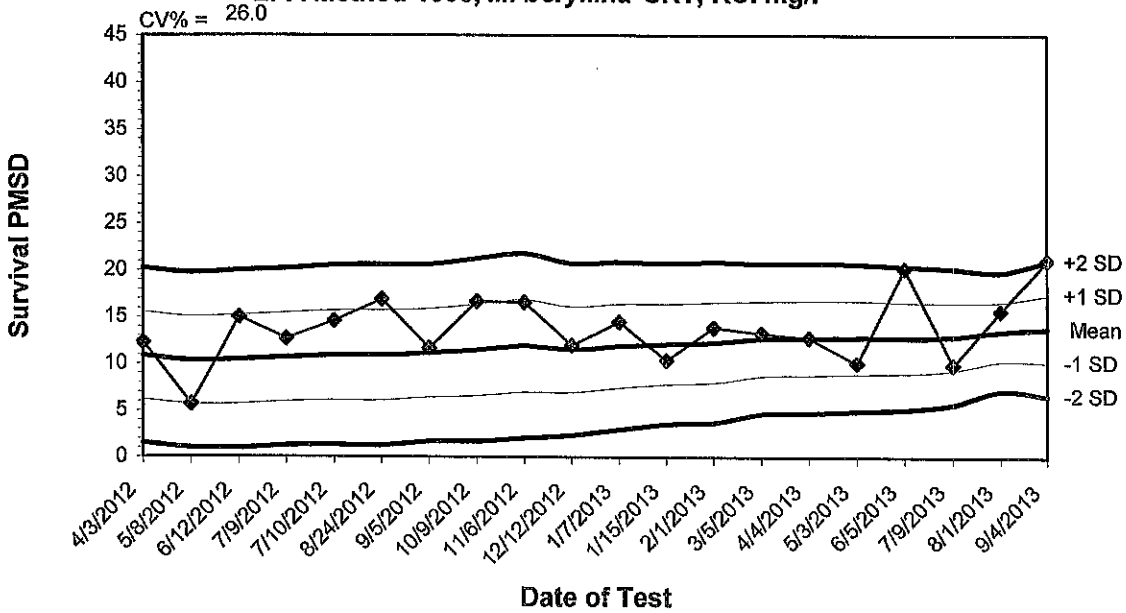
Dilution Series = 480, 686, 980, 1400, & 2000 mg/l KCl; Dilution Factor = 0.7

Test #	Test Date	Survival NOEC	% Control Survival	Survival PMSD	Upper Control Limit	Lower Control Limit	Central Tendency	SRT Lot #
MN1205	4/3/2012	980	92.5	12.3	1400	686	980	060M0116V
MN1206	5/8/2012	980	97.5	5.7	1400	686	980	021M0113V
MN1207	6/12/2012	980	95.0	15.0	1400	686	980	081M0170V
MN1208	7/9/2012	1400	95.0	12.7	1400	686	980	081M0170V
MN1209	7/10/2012	980	95.0	14.6	1400	686	980	081M0170V
MN1210	8/24/2012	980	90.0	16.9	1400	686	980	081M0170V
MN1211	9/5/2012	980	92.5	11.7	1400	686	980	081M0170V
MN1212	10/9/2012	980	97.5	16.7	1400	686	980	081M0170V
MN1213	11/6/2012	980	95.0	16.6	1400	686	980	081M0170V
MN1214	12/12/2012	980	97.5	12.0	1400	686	980	081M0170V
MN1301	1/7/2013	980	97.5	14.5	1400	686	980	081M0170V
MN1302	1/15/2013	980	97.5	10.4	1400	686	980	081M0170V
MN1303	2/1/2013	1400	100.0	13.9	1400	686	980	081M0170V
MN1304	3/5/2013	980	92.5	13.3	1400	686	980	SLBD2389V
MN1305	4/4/2013	980	95.0	12.8	1400	686	980	081M0170V
MN1306	5/3/2013	980	92.5	10.1	1400	686	980	SLBD2389V
MN1307	6/5/2013	980	95.0	20.2	1400	686	980	SLBC2414V
MN1308	7/9/2013	980	100.0	9.9	1400	686	980	SLBC2414V
MN1309	8/1/2013	980	87.5	15.7	1400	686	980	SLBC2414V
MN1310	9/4/2013	980	92.5	21.1	1400	686	980	SLBC2414V

MN1208 - Out of house organisms. Aquatic Indicators.

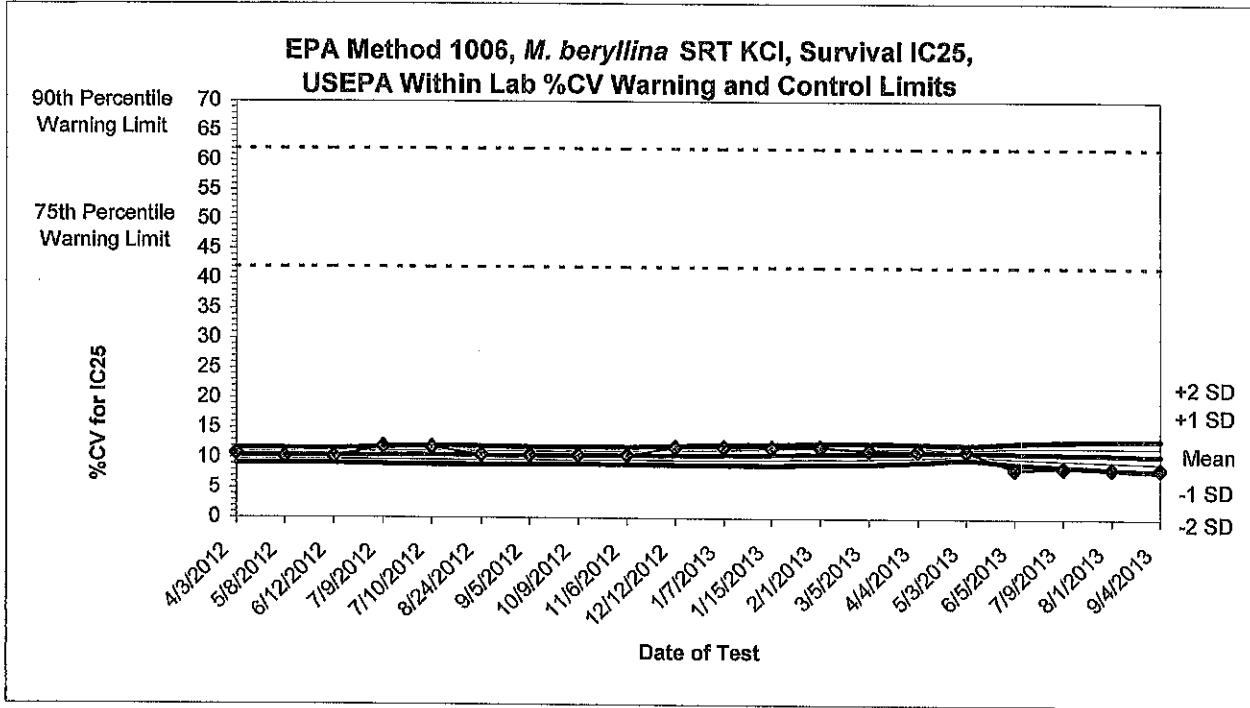
QAQC by: MAO 9/16/13

EPA Method 1006, *M. beryllina* SRT, KCl mg/l



Test #	Test Date	Survival PMSD	Mean	-1 SD	-2 SD	+1 SD	+2 SD	SRT Lot #
MN1205	4/3/2012	12.3	10.9	6.2	1.5	15.6	20.3	060M0116V
MN1206	5/8/2012	5.7	10.4	5.7	1.0	15.1	19.8	021M0113V
MN1207	6/12/2012	15.0	10.5	5.7	1.0	15.2	20.0	081M0170V
MN1208	7/9/2012	12.7	10.7	6.0	1.2	15.5	20.2	081M0170V
MN1209	7/10/2012	14.6	10.9	6.1	1.3	15.7	20.6	081M0170V
MN1210	8/24/2012	16.9	10.9	6.1	1.2	15.8	20.6	081M0170V
MN1211	9/5/2012	11.7	11.2	6.4	1.7	15.9	20.7	081M0170V
MN1212	10/9/2012	16.7	11.5	6.6	1.7	16.4	21.3	081M0170V
MN1213	11/6/2012	16.6	11.9	7.0	2.1	16.9	21.8	081M0170V
MN1214	12/12/2012	12.0	11.6	7.0	2.4	16.1	20.7	081M0170V
MN1301	1/7/2013	14.5	11.9	7.5	3.0	16.4	20.9	081M0170V
MN1302	1/15/2013	10.4	12.2	7.9	3.6	16.4	20.7	081M0170V
MN1303	2/1/2013	13.9	12.3	8.0	3.7	16.6	20.9	SLBD2389V
MN1304	3/5/2013	13.3	12.7	8.7	4.7	16.7	20.7	081M0170V
MN1305	4/4/2013	12.8	12.8	8.8	4.8	16.8	20.8	SLBD2389V
MN1306	5/3/2013	10.1	12.9	8.9	5.0	16.8	20.7	SLBD2389V
MN1307	6/5/2013	20.2	12.8	9.0	5.2	16.6	20.4	SLBC2414V
MN1308	7/9/2013	9.9	13.0	9.3	5.7	16.6	20.2	SLBC2414V
MN1309	8/1/2013	15.7	13.5	10.3	7.2	16.6	19.8	SLBC2414V
MN1310	9/4/2013	21.1	13.8	10.2	6.6	17.4	21.0	SLBC2414V

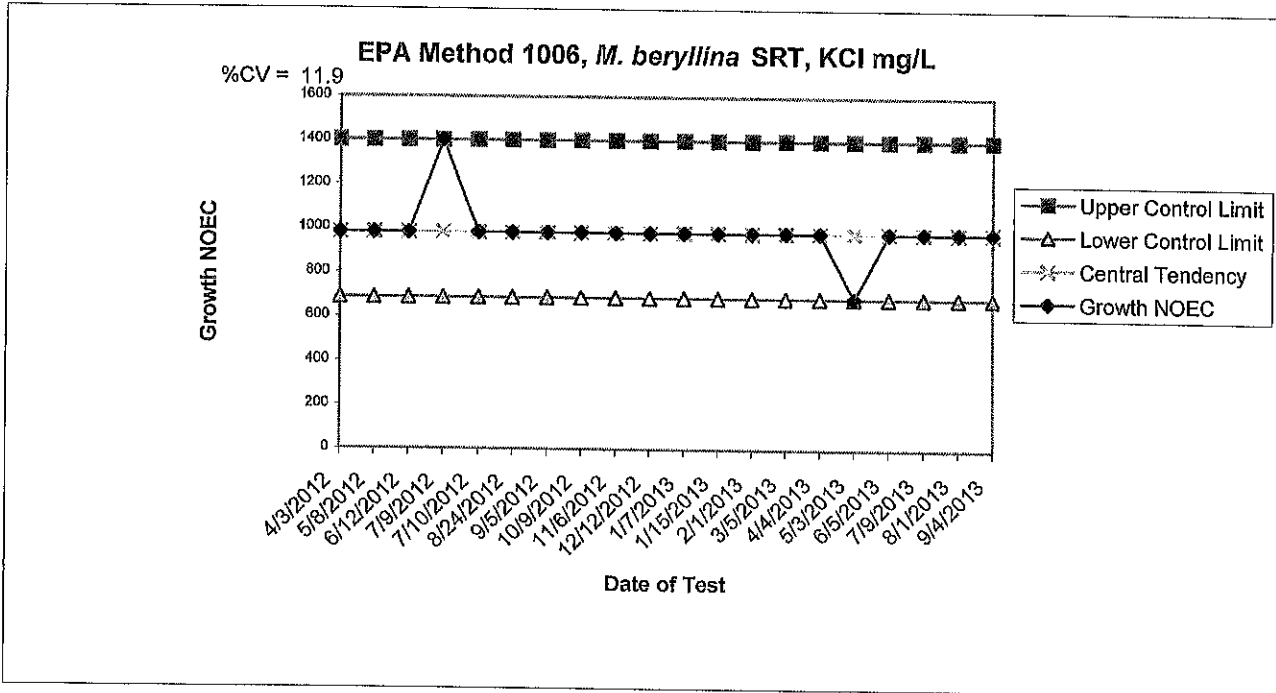
QAQC by: MMAO 9/10/13



15%CV = 10th percentile, 22%CV = 25th percentile, 35%CV = 50th percentile

Test #	Test Date	%CV for IC25	Mean %CV	-1 SD	-2 SD	+1 SD	+2 SD	75th Warning Limit	90th Warning Limit	SRT Lot #
MN1205	4/3/2012	10.8	10.4	9.8	9.1	11.1	11.7	42.0	62.0	060M0116V
MN1206	5/8/2012	10.2	10.4	9.8	9.1	11.0	11.7	42.0	62.0	021M0113V
MN1207	6/12/2012	10.2	10.4	9.7	9.1	11.0	11.6	42.0	62.0	081M0170V
MN1208	7/9/2012	11.9	10.4	9.7	9.0	11.2	11.9	42.0	62.0	081M0170V
MN1209	7/10/2012	11.8	10.5	9.7	8.9	11.3	12.0	42.0	62.0	081M0170V
MN1210	8/24/2012	10.5	10.4	9.7	8.9	11.2	12.0	42.0	62.0	081M0170V
MN1211	9/5/2012	10.5	10.4	9.7	8.9	11.2	11.9	42.0	62.0	081M0170V
MN1212	10/9/2012	10.5	10.4	9.7	8.9	11.2	11.9	42.0	62.0	081M0170V
MN1213	11/6/2012	10.5	10.4	9.7	8.9	11.2	11.9	42.0	62.0	081M0170V
MN1214	12/12/2012	11.8	10.5	9.7	8.9	11.3	12.1	42.0	62.0	081M0170V
MN1301	1/7/2013	11.9	10.6	9.7	8.9	11.4	12.3	42.0	62.0	081M0170V
MN1302	1/15/2013	11.8	10.6	9.7	8.9	11.5	12.4	42.0	62.0	081M0170V
MN1303	2/1/2013	11.9	10.8	9.9	9.0	11.7	12.5	42.0	62.0	SLBD2389V
MN1304	3/5/2013	11.4	10.8	10.0	9.1	11.7	12.6	42.0	62.0	081M0170V
MN1305	4/4/2013	11.3	11.0	10.2	9.4	11.7	12.5	42.0	62.0	SLBD2389V
MN1306	5/3/2013	11.3	11.1	10.5	9.8	11.7	12.3	42.0	62.0	SLBD2389V
MN1307	6/5/2013	8.3	11.0	10.1	9.2	11.8	12.7	42.0	62.0	SLBC2414V
MN1308	7/9/2013	8.5	10.8	9.8	8.8	11.9	12.9	42.0	62.0	SLBC2414V
MN1309	8/1/2013	8.4	10.7	9.5	8.4	11.9	13.1	42.0	62.0	SLBC2414V
MN1310	9/4/2013	8.4	10.6	9.3	8.0	11.9	13.2	42.0	62.0	SLBC2414V

QAQC by: MAO 9/16/13



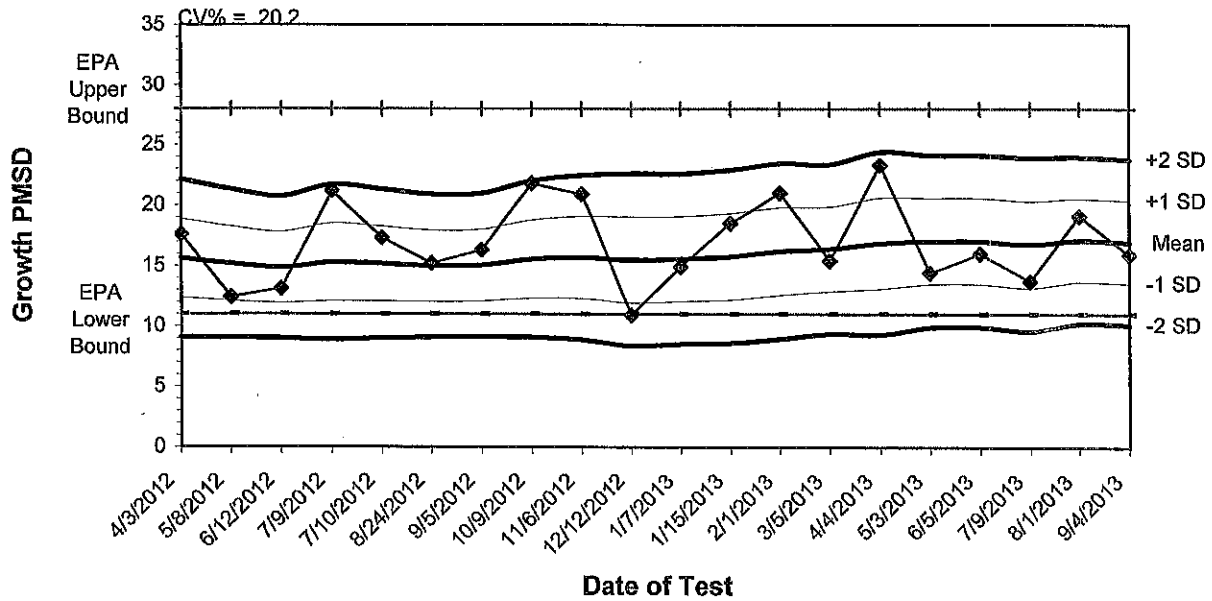
Dilution Series = 480, 686, 980, 1400, & 2000 mg/l KCl; Dilution Factor = 0.7

Test #	Test Date	Growth NOEC	Mean Control Growth	Growth PMSD	Upper Control Limit	Lower Control Limit	Central Tendency	SRT Lot #
MN1205	4/3/2012	980	1.465	17.6	1400	686	980	060M0116V
MN1206	5/8/2012	980	1.613	12.4	1400	686	980	021M0113V
MN1207	6/12/2012	980	1.557	13.1	1400	686	980	081M0170V
MN1208	7/9/2012	1400	1.526	21.2	1400	686	980	081M0170V
MN1209	7/10/2012	980	0.975	17.3	1400	686	980	081M0170V
MN1210	8/24/2012	980	1.069	15.2	1400	686	980	081M0170V
MN1211	9/5/2012	980	1.348	16.3	1400	686	980	081M0170V
MN1212	10/9/2012	980	1.122	21.8	1400	686	980	081M0170V
MN1213	11/6/2012	980	1.069	20.9	1400	686	980	081M0170V
MN1214	12/12/2012	980	1.426	10.9	1400	686	980	081M0170V
MN1301	1/7/2013	980	1.309	14.9	1400	686	980	081M0170V
MN1302	1/15/2013	980	1.165	18.5	1400	686	980	081M0170V
MN1303	2/1/2013	980	1.223	21.0	1400	686	980	SLBD2389V
MN1304	3/5/2013	980	1.153	15.4	1400	686	980	081M0170V
MN1305	4/4/2013	980	1.281	23.3	1400	686	980	SLBD2389V
MN1306	5/3/2013	686	1.616	14.4	1400	686	980	SLBD2389V
MN1307	6/5/2013	980	0.953	16.0	1400	686	980	SLBC2414V
MN1308	7/9/2013	980	1.107	13.7	1400	686	980	SLBC2414V
MN1309	8/1/2013	980	0.917	19.1	1400	686	980	SLBC2414V
MN1310	9/4/2013	980	1.013	15.9	1400	686	980	SLBC2414V

MN1208 - Out of house organisms. Aquatic Indicators.

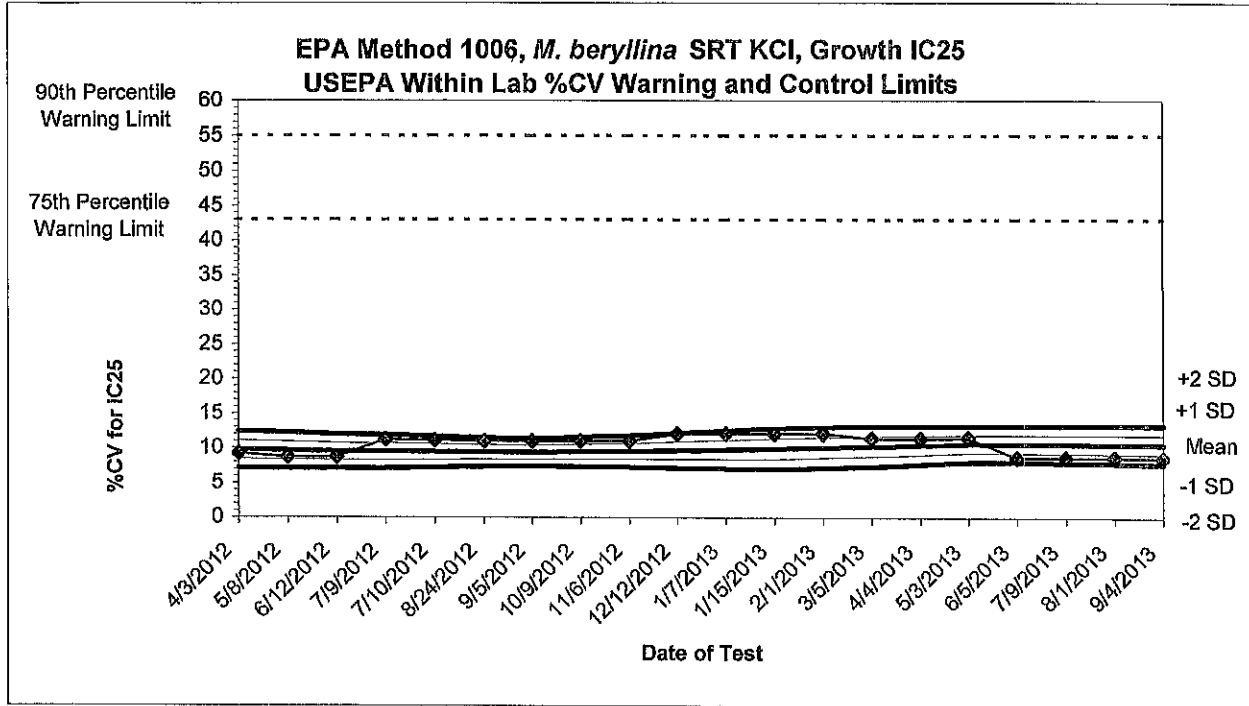
QAQC by: MNO 9/16/13

EPA Method 1006, *M. beryllina* SRT Growth PMSD



Test #	Test Date	Growth PMSD	Mean PMSD	-1 SD	-2 SD	+1 SD	+2 SD	Upper PMSD Bound	Lower PMSD Bound	SRT Lot #
MN1205	4/3/2012	17.6	15.6	12.3	9.1	18.9	22.1	28	11	060M0116V
MN1206	5/8/2012	12.4	15.2	12.1	9.0	18.3	21.3	28	11	021M0113V
MN1207	6/12/2012	13.1	14.9	11.9	9.0	17.8	20.7	28	11	081M0170V
MN1208	7/9/2012	21.2	15.3	12.1	8.9	18.5	21.7	28	11	081M0170V
MN1209	7/10/2012	17.3	15.2	12.1	9.0	18.2	21.3	28	11	081M0170V
MN1210	8/24/2012	15.2	15.0	12.0	9.1	17.9	20.9	28	11	081M0170V
MN1211	9/5/2012	16.3	15.1	12.1	9.1	18.0	21.0	28	11	081M0170V
MN1212	10/9/2012	21.8	15.5	12.3	9.1	18.8	22.0	28	11	081M0170V
MN1213	11/6/2012	20.9	15.7	12.3	8.9	19.1	22.5	28	11	081M0170V
MN1214	12/12/2012	10.9	15.5	11.9	8.4	19.0	22.6	28	11	081M0170V
MN1301	1/7/2013	14.9	15.6	12.0	8.5	19.1	22.6	28	11	081M0170V
MN1302	1/15/2013	18.5	15.7	12.2	8.6	19.3	22.9	28	11	081M0170V
MN1303	2/1/2013	21.0	16.2	12.6	8.9	19.8	23.5	28	11	SLBD2389V
MN1304	3/5/2013	15.4	16.4	12.9	9.4	19.9	23.4	28	11	081M0170V
MN1305	4/4/2013	23.3	16.8	13.1	9.3	20.6	24.4	28	11	SLBD2389V
MN1306	5/3/2013	14.4	17.0	13.5	9.9	20.6	24.1	28	11	SLBD2389V
MN1307	6/5/2013	16.0	17.0	13.5	9.9	20.6	24.1	28	11	SLBC2414V
MN1308	7/9/2013	13.7	16.7	13.2	9.6	20.3	23.9	28	11	SLBC2414V
MN1309	8/1/2013	19.1	17.1	13.7	10.2	20.5	24.0	28	11	SLBC2414V
MN1310	9/4/2013	15.9	16.9	13.5	10.1	20.4	23.8	28	11	SLBC2414V

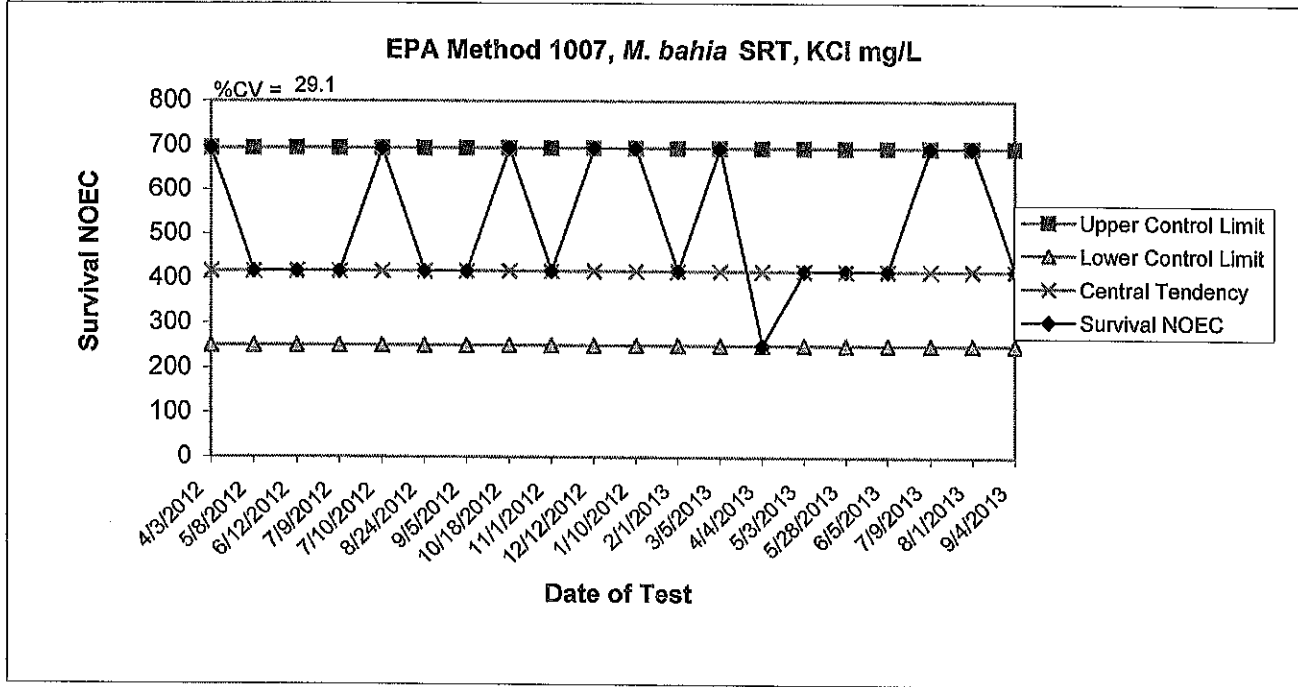
QAQC by: MAO 9/16/13



5%CV = 10th percentile, 18%CV = 25th percentile, 27%CV = 50th percentile

Test #	Test Date	%CV for IC25	Mean %CV	-1 SD	-2 SD	+1 SD	+2 SD	75th Warning Limit	90th Warning Limit	SRT Lot #
MN1205	4/3/2012	9.2	9.8	8.4	7.1	11.1	12.4	43.0	55.0	060M0116V
MN1206	5/8/2012	8.7	9.6	8.3	7.0	10.9	12.2	43.0	55.0	021M0113V
MN1207	6/12/2012	8.7	9.5	8.3	7.0	10.7	12.0	43.0	55.0	081M0170V
MN1208	7/9/2012	11.2	9.5	8.3	7.1	10.7	11.9	43.0	55.0	081M0170V
MN1209	7/10/2012	11.1	9.4	8.3	7.2	10.6	11.7	43.0	55.0	081M0170V
MN1210	8/24/2012	11.0	9.4	8.3	7.3	10.4	11.5	43.0	55.0	081M0170V
MN1211	9/5/2012	11.0	9.4	8.4	7.4	10.4	11.4	43.0	55.0	081M0170V
MN1212	10/9/2012	11.0	9.5	8.4	7.3	10.5	11.6	43.0	55.0	081M0170V
MN1213	11/6/2012	11.0	9.5	8.4	7.3	10.7	11.8	43.0	55.0	081M0170V
MN1214	12/12/2012	12.0	9.6	8.4	7.1	10.9	12.2	43.0	55.0	081M0170V
MN1301	1/7/2013	12.1	9.8	8.4	7.0	11.1	12.5	43.0	55.0	081M0170V
MN1302	1/15/2013	12.0	9.9	8.4	7.0	11.4	12.8	43.0	55.0	081M0170V
MN1303	2/1/2013	12.1	10.1	8.6	7.2	11.6	13.0	43.0	55.0	SLBD2389V
MN1304	3/5/2013	11.4	10.2	8.8	7.3	11.7	13.1	43.0	55.0	081M0170V
MN1305	4/4/2013	11.4	10.4	9.0	7.7	11.8	13.2	43.0	55.0	SLBD2389V
MN1306	5/3/2013	11.5	10.6	9.3	8.0	11.9	13.1	43.0	55.0	SLBD2389V
MN1307	6/5/2013	8.6	10.6	9.3	8.0	11.9	13.2	43.0	55.0	SLBC2414V
MN1308	7/9/2013	8.6	10.5	9.2	7.9	11.9	13.2	43.0	55.0	SLBC2414V
MN1309	8/1/2013	8.7	10.5	9.2	7.8	11.9	13.2	43.0	55.0	SLBC2414V
MN1310	9/4/2013	8.6	10.5	9.1	7.7	11.9	13.3	43.0	55.0	SLBC2414V

QAQC by: MAO 9/10/13



Dilution Series = 150, 250, 416, 694, & 1157 mg/l KCl; Dilution Factor = 0.60

Test #	Test Date	Survival NOEC	% Control Survival	Survival PMSD	Upper Control Limit	Lower Control Limit	Central Tendency	SRT Lot #
MB1204	4/3/2012	694	97.5	9.0	694	250	416	060M0116V
MB1205	5/8/2012	416	97.5	7.4	694	250	416	021M0113V
MB1206	6/12/2012	416	97.5	14.3	694	250	416	081M0170V
MB1208	7/9/2012	416	95.0	8.1	694	250	416	081M0170V
MB1209	7/10/2012	694	100.0	5.1	694	250	416	081M0170V
MB1210	8/24/2012	416	100.0	7.0	694	250	416	081M0170V
MB1211	9/5/2012	416	95.0	15.8	694	250	416	081M0170V
MB1212	10/18/2012	694	97.5	10.5	694	250	416	081M0170V
MB1213	11/1/2012	416	100.0	7.5	694	250	416	081M0170V
MB1214	12/12/2012	694	100.0	13.9	694	250	416	081M0170V
MB1301	1/10/2012	694	100.0	4.5	694	250	416	081M0170V
MB1302	2/1/2013	416	100.0	4.6	694	250	416	SLBD2389V
MB1303	3/5/2013	694	100.0	10.7	694	250	416	081M0170V
MB1304	4/4/2013	250	100.0	6.5	694	250	416	SLBD2389V
MB1305	5/3/2013	416	100.0	12.6	694	250	416	SLBD2389V
MB1306	5/28/2013	416	97.5	8.1	694	250	416	SLBD2389V
MB1307	6/5/2013	416	94.3	17.0	694	250	416	SLBD2389V
MB1308	7/9/2013	694	100.0	5.9	694	250	416	SLBC2414V
MB1309	8/1/2013	694	100.0	12.6	694	250	416	SLBC2414V
MB1310	9/4/2013	416	100.0	7.3	694	250	416	SLBC2414V

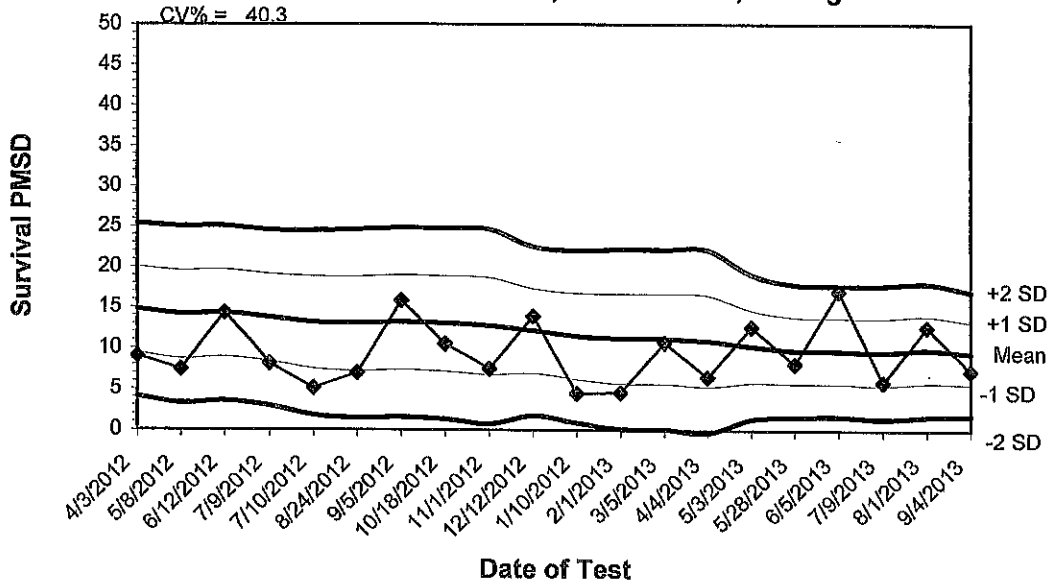
MB1207 - Number was not used.

MB1208 - Out of house organisms. Marinc Bioassay Laboratories.

MB1307 - Lot# SLBD2389V was used to mix Day 0, Day 1 - 6 was mixed with Lot# SLBC2414V.

QAQC by: MAO 9/17/13

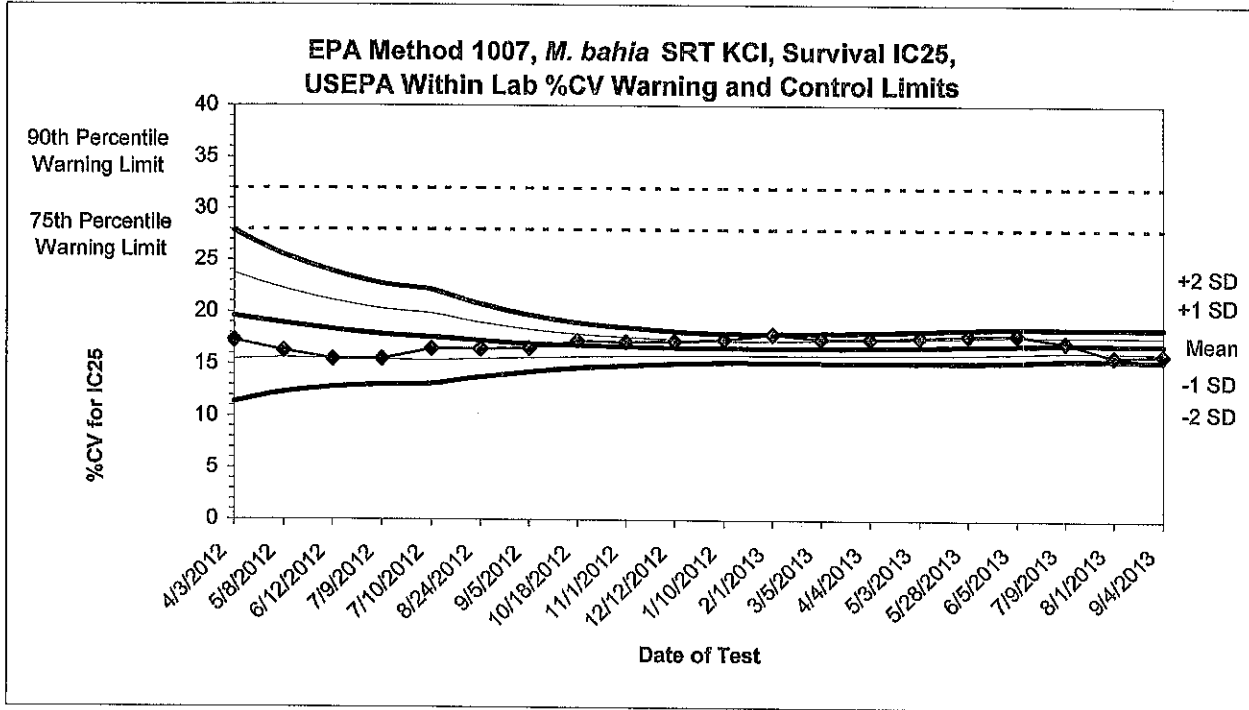
EPA Method 1007, *M. bahia* SRT, KCl mg/l



Test #	Test Date	Survival PMSD	Mean PMSD	-1 SD	-2 SD	+1 SD	+2 SD	SRT Lot #
MB1204	4/3/2012	9.0	14.8	9.4	4.1	20.1	25.4	060M0116V
MB1205	5/8/2012	7.4	14.2	8.7	3.3	19.6	25.0	021M0113V
MB1206	6/12/2012	14.3	14.3	8.9	3.5	19.7	25.1	081M0170V
MB1208	7/9/2012	8.1	13.7	8.3	2.9	19.1	24.5	081M0170V
MB1209	7/10/2012	5.1	13.1	7.5	1.8	18.8	24.5	081M0170V
MB1210	8/24/2012	7.0	13.0	7.2	1.4	18.8	24.6	081M0170V
MB1211	9/5/2012	15.8	13.2	7.4	1.5	19.0	24.8	081M0170V
MB1212	10/18/2012	10.5	13.0	7.2	1.3	18.9	24.7	081M0170V
MB1213	11/1/2012	7.5	12.7	6.7	0.8	18.7	24.7	081M0170V
MB1214	12/12/2012	13.9	12.1	6.9	1.8	17.3	22.5	081M0170V
MB1301	1/10/2012	4.5	11.5	6.2	0.9	16.8	22.1	081M0170V
MB1302	2/1/2013	4.6	11.2	5.7	0.2	16.7	22.2	SLBD2389V
MB1303	3/5/2013	10.7	11.1	5.6	0.1	16.7	22.2	081M0170V
MB1304	4/4/2013	6.5	10.9	5.3	-0.3	16.5	22.1	SLBD2389V
MB1305	5/3/2013	12.6	10.2	5.8	1.4	14.7	19.1	SLBD2389V
MB1306	5/28/2013	8.1	9.7	5.7	1.6	13.8	17.8	SLBD2389V
MB1307	6/5/2013	17.0	9.7	5.7	1.7	13.7	17.7	SLBD2389V
MB1308	7/9/2013	5.9	9.6	5.5	1.4	13.7	17.8	SLBC2414V
MB1309	8/1/2013	12.6	9.9	5.8	1.7	14.0	18.0	SLBC2414V
MB1310	9/4/2013	7.3	9.4	5.6	1.8	13.2	17.0	SLBC2414V

MB1307 - Lot# SLBD2389V was used to mix Day 0, Day 1 - 6 was mixed with Lot# SLBC2414V.

QAQC by: MA 9/17/13

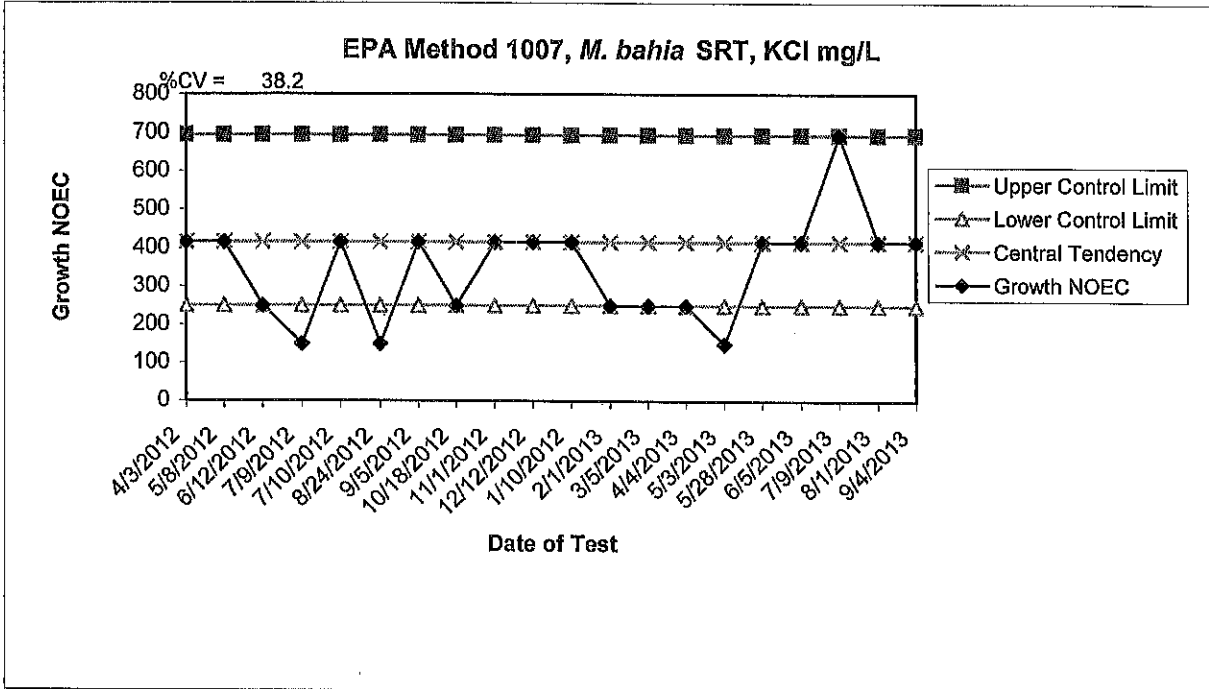


17%CV = 10th percentile, 17%CV = 25th percentile, 21%CV = 50th percentile

Test #	Test Date	%CV for IC25	Mean %CV	-1 SD	-2 SD	+1 SD	+2 SD	75th Warning Limit	90th Warning Limit	SRT Lot #
MB1204	4/3/2012	17.3	19.6	15.5	11.4	23.7	27.9	28.0	32.0	060M0116V
MB1205	5/8/2012	16.3	18.9	15.6	12.3	22.2	25.5	28.0	32.0	021M0113V
MB1206	6/12/2012	15.5	18.3	15.6	12.8	21.1	23.9	28.0	32.0	081M0170V
MB1208	7/9/2012	15.5	17.9	15.5	13.0	20.3	22.7	28.0	32.0	081M0170V
MB1209	7/10/2012	16.5	17.6	15.4	13.1	19.8	22.1	28.0	32.0	081M0170V
MB1210	8/24/2012	16.5	17.2	15.5	13.8	19.0	20.7	28.0	32.0	081M0170V
MB1211	9/5/2012	16.6	17.0	15.6	14.3	18.3	19.7	28.0	32.0	081M0170V
MB1212	10/18/2012	17.3	16.8	15.7	14.6	17.9	19.0	28.0	32.0	081M0170V
MB1213	11/1/2012	17.2	16.7	15.8	14.9	17.6	18.6	28.0	32.0	081M0170V
MB1214	12/12/2012	17.2	16.6	15.8	15.1	17.4	18.2	28.0	32.0	081M0170V
MB1301	1/10/2012	17.4	16.6	15.9	15.2	17.3	18.0	28.0	32.0	081M0170V
MB1302	2/1/2013	17.9	16.6	15.9	15.2	17.3	18.0	28.0	32.0	SLBD2389V
MB1303	3/5/2013	17.5	16.6	15.9	15.2	17.3	18.0	28.0	32.0	081M0170V
MB1304	4/4/2013	17.5	16.6	15.9	15.2	17.4	18.1	28.0	32.0	SLBD2389V
MB1305	5/3/2013	17.6	16.7	15.9	15.2	17.5	18.2	28.0	32.0	SLBD2389V
MB1306	5/28/2013	17.8	16.8	16.0	15.2	17.6	18.4	28.0	32.0	SLBD2389V
MB1307	6/5/2013	17.9	16.9	16.1	15.3	17.7	18.5	28.0	32.0	SLBD2389V
MB1308	7/9/2013	17.2	17.0	16.2	15.5	17.7	18.5	28.0	32.0	SLBC2414V
MB1309	8/1/2013	15.8	17.0	16.2	15.5	17.7	18.5	28.0	32.0	SLBC2414V
MB1310	9/4/2013	16.0	16.9	16.1	15.3	17.7	18.5	28.0	32.0	SLBC2414V

MB1307 - Lot# SLBD2389V was used to mix Day 0, Day 1 - 6 was mixed with Lot# SLBC2414V.

QAQC by: MAO 9/17/13



Dilution Series = 150, 250, 416, 694, & 1157 mg/l KCl; Dilution Factor = 0.60

Test #	Test Date	Growth NOEC	Mean Control Growth	Growth PMSD	Upper Control Limit	Lower Control Limit	Central Tendency	SRT Lot #
MB1204	4/3/2012	416	0.335	18.1	694	250	416	060M0116V
MB1205	5/8/2012	416	0.329	17.1	694	250	416	021M0113V
MB1206	6/12/2012	250	0.347	13.1	694	250	416	081M0170V
MB1208	7/9/2012	150	0.239	13.2	694	250	416	081M0170V
MB1209	7/10/2012	416	0.369	18.2	694	250	416	081M0170V
MB1210	8/24/2012	150	0.212	12.8	694	250	416	081M0170V
MB1211	9/5/2012	416	0.364	16.1	694	250	416	081M0170V
MB1212	10/18/2012	250	0.343	13.2	694	250	416	081M0170V
MB1213	11/1/2012	416	0.312	11.5	694	250	416	081M0170V
MB1214	12/12/2012	416	0.321	21.8	694	250	416	081M0170V
MB1301	1/10/2012	416	0.230	14.8	694	250	416	081M0170V
MB1302	2/1/2013	250	0.376	8.1	694	250	416	SLBD2389V
MB1303	3/5/2013	250	0.326	17.0	694	250	416	081M0170V
MB1304	4/4/2013	250	0.307	12.6	694	250	416	SLBD2389V
MB1305	5/3/2013	150	0.318	9.4	694	250	416	SLBD2389V
MB1306	5/28/2013	416	0.301	14.7	694	250	416	SLBD2389V
MB1307	6/5/2013	416	0.309	14.4	694	250	416	SLBD2389V
MB1308	7/9/2013	694	0.277	25.3	694	250	416	SLBC2414V
MB1309	8/1/2013	416	0.296	18.7	694	250	416	SLBC2414V
MB1310	9/4/2013	416	0.371	13.2	694	250	416	SLBC2414V

MB1207 - Number was not used.

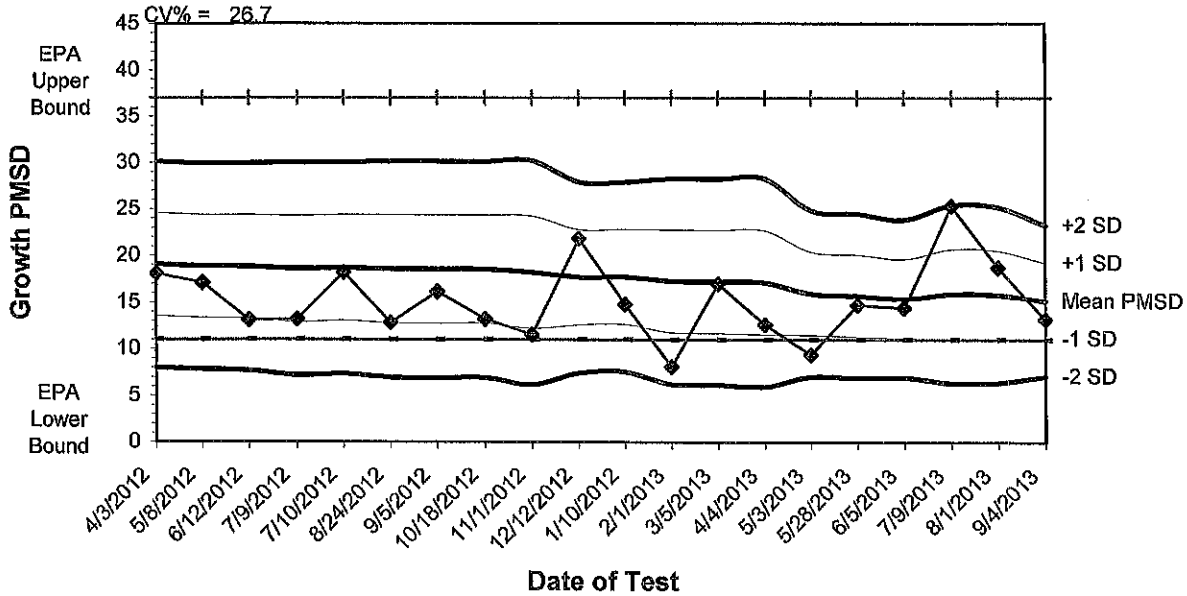
MB1208 - Out of house organisms. Marinc Bioassay Laboratories. This is a valid result.

MB1305 - Growth NOEC was less than the lower control limit. An additional test will be completed in May.

MB1307 - Lot# SLBD2389V was used to mix Day 0, Day 1 - 6 was mixed with Lot# SLBC2414V.

QAQC by: MAO 9/17/13

EPA Method 1007, *M. bahia* SRT Growth PMSD

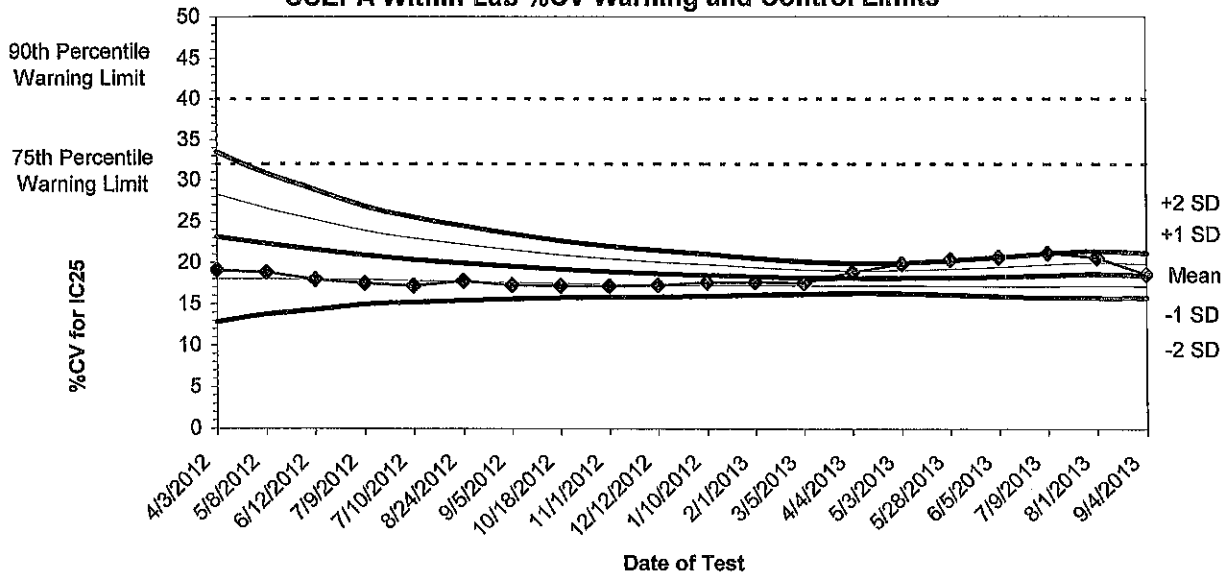


Test #	Test Date	Growth PMSD	Mean PMSD	-1 SD	-2 SD	+1 SD	+2 SD	Upper PMSD Bound	Lower PMSD Bound	SRT Lot #
MB1204	4/3/2012	18.1	19.1	13.5	8.0	24.6	30.1	37	11	060M0116V
MB1205	5/8/2012	17.1	18.9	13.4	7.8	24.4	29.9	37	11	021M0113V
MB1206	6/12/2012	13.1	18.8	13.3	7.7	24.4	30.0	37	11	081M0170V
MB1208	7/9/2012	13.2	18.6	12.9	7.2	24.3	30.0	37	11	081M0170V
MB1209	7/10/2012	18.2	18.7	13.0	7.4	24.4	30.1	37	11	081M0170V
MB1210	8/24/2012	12.8	18.6	12.8	7.0	24.4	30.1	37	11	081M0170V
MB1211	9/5/2012	16.1	18.5	12.7	6.9	24.3	30.1	37	11	081M0170V
MB1212	10/18/2012	13.2	18.5	12.7	6.9	24.3	30.1	37	11	081M0170V
MB1213	11/1/2012	11.5	18.2	12.2	6.2	24.2	30.2	37	11	081M0170V
MB1214	12/12/2012	21.8	17.7	12.6	7.4	22.8	27.9	37	11	081M0170V
MB1301	1/10/2013	14.8	17.7	12.7	7.6	22.8	27.9	37	11	081M0170V
MB1302	2/1/2013	8.1	17.3	11.7	6.2	22.8	28.3	37	11	SLBD2389V
MB1303	3/5/2013	17.0	17.2	11.7	6.2	22.7	28.2	37	11	081M0170V
MB1304	4/4/2013	12.6	17.1	11.5	6.0	22.7	28.3	37	11	SLBD2389V
MB1305	5/3/2013	9.4	15.9	11.5	7.0	20.4	24.9	37	11	SLBD2389V
MB1306	5/28/2013	14.7	15.7	11.3	6.9	20.1	24.5	37	11	SLBD2389V
MB1307	6/5/2013	14.4	15.4	11.2	6.9	19.6	23.8	37	11	SLBD2389V
MB1308	7/9/2013	25.3	15.9	11.1	6.4	20.7	25.4	37	11	SLBC2414V
MB1309	8/1/2013	18.7	15.8	11.1	6.4	20.5	25.2	37	11	SLBC2414V
MB1310	9/4/2013	13.2	15.2	11.1	7.1	19.2	23.3	37	11	SLBC2414V

MB1307 - Lot# SLBD2389V was used to mix Day 0, Day 1 - 6 was mixed with Lot# SLBC2414V.

QAQC by: MAO 9/17/13

**EPA Method 1007, *M. bahia* SRT KCI, Growth IC25,
USEPA Within Lab %CV Warning and Control Limits**



21%CV = 10th percentile, 24%CV = 25th percentile, 28%CV = 50th percentile

Test #	Test Date	%CV for IC25	Mean %CV	-1 SD	-2 SD	+1 SD	+2 SD	75th Warning Limit	90th Warning Limit	SRT Lot #
MB1204	4/3/2012	19.1	23.2	18.0	12.8	28.3	33.5	32.0	40.0	060M0116V
MB1205	5/8/2012	18.8	22.3	18.0	13.8	26.6	30.8	32.0	40.0	021M0113V
MB1206	6/12/2012	17.9	21.6	17.9	14.3	25.2	28.8	32.0	40.0	081M0170V
MB1208	7/9/2012	17.5	20.9	17.9	14.9	23.9	26.8	32.0	40.0	081M0170V
MB1209	7/10/2012	17.2	20.4	17.8	15.2	22.9	25.5	32.0	40.0	081M0170V
MB1210	8/24/2012	17.7	19.9	17.7	15.4	22.2	24.5	32.0	40.0	081M0170V
MB1211	9/5/2012	17.2	19.5	17.6	15.6	21.5	23.5	32.0	40.0	081M0170V
MB1212	10/18/2012	17.2	19.2	17.5	15.7	20.9	22.6	32.0	40.0	081M0170V
MB1213	11/1/2012	17.1	18.9	17.4	15.8	20.4	22.0	32.0	40.0	081M0170V
MB1214	12/12/2012	17.2	18.7	17.3	15.9	20.1	21.5	32.0	40.0	081M0170V
MB1301	1/10/2012	17.6	18.5	17.2	15.9	19.8	21.0	32.0	40.0	081M0170V
MB1302	2/1/2013	17.6	18.3	17.2	16.1	19.4	20.5	32.0	40.0	SLBD2389V
MB1303	3/5/2013	17.5	18.1	17.2	16.2	19.1	20.1	32.0	40.0	081M0170V
MB1304	4/4/2013	18.8	18.1	17.2	16.3	19.0	19.9	32.0	40.0	SLBD2389V
MB1305	5/3/2013	19.8	18.1	17.2	16.2	19.0	20.0	32.0	40.0	SLBD2389V
MB1306	5/28/2013	20.4	18.2	17.1	16.1	19.2	20.3	32.0	40.0	SLBD2389V
MB1307	6/5/2013	20.7	18.3	17.1	15.9	19.5	20.7	32.0	40.0	SLBD2389V
MB1308	7/9/2013	21.1	18.4	17.1	15.7	19.8	21.1	32.0	40.0	SLBC2414V
MB1309	8/1/2013	20.6	18.6	17.1	15.7	20.0	21.4	32.0	40.0	SLBC2414V
MB1310	9/4/2013	18.6	18.5	17.1	15.8	19.8	21.2	32.0	40.0	SLBC2414V

MB1307 - Lot# SLBD2389V was used to mix Day 0, Day 1 - 6 was mixed with Lot# SLBC2414V.

QAQC by: NAO 9/17/13

Environmental Enterprises USA, Inc.

APPENDIX G

CorrLine International – CorrX
Run-off from treated metal plate
 Barbara Tompkins-Brown

Test Concentrations, % Prepared Chemically Treated Seawater (CTS)

<i>Menidia beryllina</i>	<i>Mysidopsis bahia</i>	Total Volume/ Concentration, ml	Color Code	ml CTS	ml DH ₂ O
0.30		800.00	Black	2.40	797.60
0.10		"	Red	0.80	799.20
0.03		"	Yellow	0.24	799.76
0.01		"	Green	0.08	799.92
0.003		"	Blue	0.02	799.98
0.0		"	White	0.00	800.00
Total Volume (ml) of CTS needed per day=					3.54
Total Volume (ml) of CTS needed for test duration=					7.08

Data Pages & Calculations by: *Kerence M. [Signature]* QA/QC Check by: *Nicole [Signature]*

M. beryllina = 2 Reps x 200 ml
= 400 ml

M. bahia = 2 Reps x 200 ml
= 400 ml

DH₂O = Dilution Water = **Synthetic Seawater, 25 ppt**

	LPC	M #	LPC	M #
Date	8/29		8/31	
Alkalinity	108	//	104	//
Salinity	24.8	1B	24.7	1B
pH	8.0	AFB	8.0	AFB
	duj		ME	

Artemia Lot #	
042012-2	
Initial	ME

LPC: Laboratory Performance Control, synthetic seawater
 Alkalinity: mg/l as CaCO₃ Salinity: ppt pH: su M#: meter number

Prep Date	8/29	8/31
DH ₂ O Lot #	25R- 239 -13	25R- 240 -13
Sample #	1	1
Initial	CM	JC

Inland silverside Minnow, *Menidia beryllina*

Acute Static-Renewal 48-Hour Definitive Test

EPA-821-R-02-012: Section 9 Method 2006

**CorrLine International – CorrX
Run-off from treated metal plate**

Test Organisms Age: 12 Days Old Test Organisms Source: EE

Test Initiation At: 1711 on 8/29/13

Counted by: Judy Merien QC/QA by: Judy Merien
Loaded by: Judy Merien Organism Lot #: mn-227-13

Exposure Chamber: 16 oz. plastic cups. Feeding: None.

Survival Data

Treatment % CTS

Time	REP	LPC White	REP	6.25 Blue	REP	12.5 Green	REP	25.0 Yellow	REP	50.0 Red	REP	100.0 Black	Initials
0 HR <u>1711</u>	1	8	3	8	5	8	7	8	9	8	11	8	8/29/13 <u>JAG</u>
	2	8	4	8	6	8	8	8	10	8	12	8	
		///		///		///		///		///		///	
24 HR <u>1024</u>	1	8	3	8	5	8	7	8	9	0	11	0	8/30/13 <u>JA</u>
	2	8	4	8	6	8	8	8	10	0	12	0	
		///		///		///		///		///		///	
48 HR <u>1310</u>	1	8	3	8	5	8	7	8	9	0	11	0	8/31/13 <u>CMB</u>
	2	8	4	8	6	8	8	8	10	0	12	0	
		///		///		///		///		///		///	
72 HR <u>1114</u>	1	8	3	8	5	8	7	8	9	0	11	0	9/1/13 <u>JA</u>
	2	8	4	8	6	8	8	8	10	0	12	0	
		///		///		///		///		///		///	
96 HR <u>1511</u>	1	8	3	8	5	8	7	8	9	0	11	0	9/2/13 <u>CA</u>
	2	8	4	8	6	8	8	8	10	0	12	0	
		///		///		///		///		///		///	
% Surv		<u>100</u>		<u>100</u>		<u>100</u>		<u>100</u>		<u>0</u>		<u>0</u>	

Data Entry by: Veronica McNew
Double Data Entry: Veronica McNew or
QA/QC Officer: Man A. O. King

M. beryllina Water Quality Data

All Treatments: Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

0 HR	Treatment % CTS						
8/29/13	LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO I	7.3	7.2	7.2	7.1	7.2	7.3	57
Temp I	24.2	24.0	24.8	25.8	24.3	23.8	1B
Salinity I	24.8	24.6	24.0	23.7	22.6	20.9	1B
Tech Initials: TK				Time: 1705			

Comments _____

24 HR	Treatment % CTS						
8/30/13	LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO F	5.8	4.4	3.6	3.3	6.3	6.3	57
Temp F	25.6	25.0	25.5	25.0	25.4	25.1	1B
Salinity F	25.1	24.9	24.6	24.1	23.3	21.5	1B
pH F	7.7	7.3	7.1	7.7	7.5	4.2	1193
Tech Initials: CJCMB				Time: 0946			

Comments Initiated aeration
at 8lml/min on
Day 1. 08/30/13 1/2

48 HR	Treatment % CTS						
8/31/13	LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO F	6.6	6.6	6.0	5.9			57
Temp F	25.3 ^{26.4}	26.3	25.7	26.2			1B
Salinity F	26.0 ^{25.3}	25.1	25.3	24.3			1B
pH F	8.0	8.0	7.8	7.6			1193
Tech Initials: CMB				Time: 0811			

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

M. beryllina Water Quality Data

All Treatments: Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

48 HR		Treatment % CTS						
8/31/13		LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO	I	7.1	7.4	7.4	7.2			S7
Temp	I	24.7	24.3	24.7	24.6			IB
Salinity	I	24.7	26.9	27.2	26.2			IB
Tech Initials: CMB		Time: 1113						

Comments _____

72 HR (A)		Treatment % CTS						
9/1/13		LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO	F							
Temp	F							
Salinity	F							
pH	F							
Tech Initials:		Time:						

Comments data not recorded
8/2/13 v2

96 HR		Treatment % CTS						
9/2/13		LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO	F	6.9	6.9	3.1	2.9			S7
Temp	F	26.5	26.5	26.3	26.4			IB
Salinity	F	25.3	27.5	27.4	27.1			IB
pH	F	8.0	8.2	7.8	7.7			A93
Tech Initials: CMB ME		Time: 0822						

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

Mysid, Mysidopsis bahia
 Acute Static-Renewal 48-Hour Definitive Test
 EPA-821-R-02-012: Section 9 Method 2007

CorrLine International – CorrX Run-off from treated metal plate

Test Organisms Age: 5 Days Old Test Organisms Source: EE
 Test Initiation At: 1113 on 08/29/13
 Counted by: Veronica McNew QC/QA by: Veronica McNew
 Loaded by: Veronica McNew Organism Lot #: MB-361-13

Exposure Chamber: 16 oz. plastic cups. Feeding: None.

Survival Data

Treatment % CTS													
Time	REP	LPC White	REP	6.25 Blue	REP	12.5 Green	REP	25.0 Yellow	REP	50.0 Red	REP	100.0 Black	Initials
0 HR <u>1113</u>	1	8	3	8	5	8	7	8	9	8	11	8	8/29/13 <u>VM</u>
	2	8	4	8	6	8	8	8	10	8	12	8	
		///		///		///		///		///		///	
24 HR <u>1020</u>	1	8	3	8	5	8	7	2	9	0	11	0	8/30/13 <u>VM</u>
	2	8	4	8	6	7	8	2	10	0	12	0	
		///		///		///		///		///		///	
48 HR <u>1320</u>	1	8	3	6	5	^(A) 23	7	0	9	0	11	0	8/31/13 <u>CMB</u>
	2	8	4	8	6	4	8	0	10	0	12	0	
		///		///		///		///		///		///	
72 HR <u>1117</u>	1	8	3	2	5	0	7	0	9	0	11	0	9/1/13 <u>JP</u>
	2	8	4	1	6	0	8	0	10	0	12	0	
		///		///		///		///		///		///	
96 HR <u>1513</u>	1	8	3	0	5	0	7	0	9	0	11	0	9/2/13 <u>CH</u>
	2	7	4	0	6	0	8	0	10	0	12	0	
		///		///		///		///		///		///	
% Surv		<u>93.8</u>		<u>0</u>		<u>0</u>		<u>0</u>		<u>0</u>		<u>0</u>	

Data Entry by: Veronica McNew
 Double Data Entry: Veronica McNew or
 QA/QC Officer: Man A. O'Neil

(A) wrong data 08-31-13 CMB

M. bahia Water Quality Data

All Treatments: Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

0 HR	Treatment % CTS						
8/29/13	LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO I	7.3	7.2	7.2	7.1	7.2	7.3	S7
Temp I	24.2	24.0	24.8	25.8	24.3	23.8	1B
Salinity I	24.8	24.4	24.0	23.7	22.6	20.9	1B
Tech Initials: TK				Time: 1705			

Comments _____

24 HR	Treatment % CTS						
8/30/13	LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO F	5.1	3.2	2.6	3.0	6.5	6.1	S7
Temp F	24.9	25.7	25.0	25.5	25.2	25.7	1B
Salinity F	25.1	24.9	24.6	24.0	23.1	21.2	1B
pH F	7.6	7.2	7.0	7.6	4.4	4.2	A93
Tech Initials: CJCMB				Time: 0949			

Comments _____
 Initiated aeration
 at 8l/min on
 Day 1. 08/30/13 VZ

48 HR	Treatment % CTS						
8/31/13	LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO F	6.8	6.7	6.5	6.4			S7
Temp F	25.6	26.3	26.3	26.3			1B
Salinity F	26.0	25.1	24.7	24.2			1B
pH F	8.0	8.0	7.9	7.7			A93
Tech Initials: CMB				Time: 0813			

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

M. bahia Water Quality Data

All Treatments: Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

48 HR	Treatment % CTS						
8/31/13	LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO I	7.1	7.4	7.4	7.2			S7
Temp I	24.7	24.3	24.7	24.6			1B
Salinity I	24.7	26.9	27.2	26.2			1B
Tech Initials: <i>CM13</i>				Time: <i>1113</i>			

Comments _____

72 HR	Treatment % CTS						
9/1/13 (A)	LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO F							
Temp F							
Salinity F							
pH F							
Tech Initials:				Time:			

Comments *(A) data not recorded*
29/02/13 v2

96 HR	Treatment % CTS						
9/2/13	LPC	6.25	12.5	25.0	50.0	100.0	Meter #
DO F	6.9	6.8					S7
Temp F	25.6	26.4					1B
Salinity F	25.8	27.2					1B
pH F	8.0	8.2					A93
Tech Initials: <i>MFCMB</i>				Time: <i>0821</i>			

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

Acute Toxicity Test-96 Hr Survival

Start Date: 8/29/2013	Test ID: mn58213	Sample ID: LAB-Lab Sample
End Date: 9/2/2013	Lab ID: EE-Environmental Enterprise	Sample Type: PRD-Product
Sample Date:	Protocol: EPAM 02-EPA Marine	Test Species: MB-Menidia beryllina
Comments:		

Conc-%	1	2
PC-LP Control	1.0000	1.0000
6.25	1.0000	1.0000
12.5	1.0000	1.0000
25	1.0000	1.0000
50	0.0000	0.0000
100	0.0000	0.0000

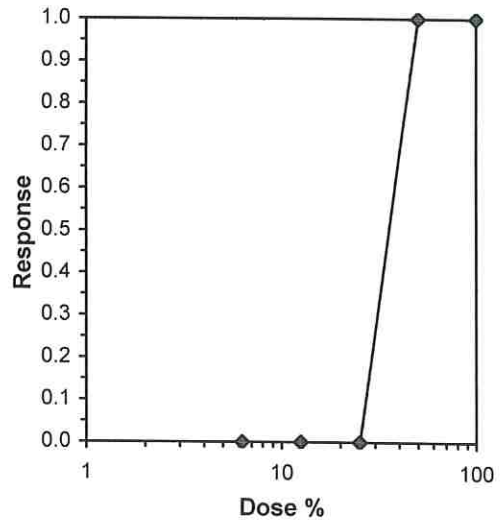
Conc-%	Mean	N-Mean	Transform: Untransformed					N	Number Resp	Total Number
			Mean	Min	Max	CV%				
PC-LP Control	1.0000	1.0000	1.0000	1.0000	1.0000	0.000	2	0	16	
6.25	1.0000	1.0000	1.0000	1.0000	1.0000	0.000	2	0	16	
12.5	1.0000	1.0000	1.0000	1.0000	1.0000	0.000	2	0	16	
25	1.0000	1.0000	1.0000	1.0000	1.0000	0.000	2	0	16	
50	0.0000	0.0000	0.0000	0.0000	0.0000	0.000	2	16	16	
100	0.0000	0.0000	0.0000	0.0000	0.0000	0.000	2	16	16	

Auxiliary Tests	Statistic	Critical	Skew	Kurt
Normality of the data set cannot be confirmed				
Equality of variance cannot be confirmed				

Graphical Method

Trim Level	EC50
0.0%	35.355

35.355



Acute Toxicity Test-96 Hr Survival

Start Date: 8/29/2013	Test ID: mb58213	Sample ID: LAB-Lab Sample
End Date: 9/2/2013	Lab ID: EE-Environmental Enterprise	Sample Type: PRD-Product
Sample Date:	Protocol: EPAM 02-EPA Marine	Test Species: MY-Mysidopsis bahia
Comments:		

Conc-%	1	2
PC-LP Control	1.0000	0.8750
6.25	0.0000	0.0000
12.5	0.0000	0.0000
25	0.0000	0.0000
50	0.0000	0.0000
100	0.0000	0.0000

Transform: Untransformed

Conc-%	Mean	N-Mean	Mean	Min	Max	CV%	N
--------	------	--------	------	-----	-----	-----	---

CorrLine International – CorrX
Run-off from treated metal plate
 Barbara Tompkins-Brown

Test Concentrations, % Prepared Chemically Treated Seawater (CTS)

	<i>Mysidopsis bahia</i>	Total Volume/ Concentration, ml	Color Code	ml CTS	ml DH ₂ O
	8.00	400.00	Black	32.00	368.00
	4.00	"	Red	16.00	374.00
	2.00	"	Yellow	8.00	392.00
	1.00	"	Green	4.00	396.00
	0.10	"	Blue	0.40	399.60
	0.0	"	White	0.00	400.00
Total Volume (ml) of CTS needed per day=					60.40
Total Volume (ml) of CTS needed for test duration=					120.80

Data Pages & Calculations by: Veronica M. [Signature] QA/QC Check by: Miranda Robbins

M. bahia = 2 Reps x 200 ml
 = 400 ml

DH₂O = Dilution Water = **Synthetic Seawater, 25 ppt**

	LPC	M #	LPC	M #
Date	9/3		9/5	
Alkalinity	104	//	106	//
Salinity	249	15	245	16
pH	8.0	A93	8.0	A93
	CMB		CMB	

Artemia Lot #	
042012-2	
Initial	MR

LPC: Laboratory Performance Control, synthetic seawater
 Alkalinity: mg/l as CaCO₃ Salinity: ppt pH: su M#: meter number

Prep Date	9/3	9/5
DH ₂ O Lot #	25R- 244 -13	25R- 246 -13
Sample #	1	1
Initial	ay	CMB

Mysid, *Mysidopsis bahia*

Acute Static-Renewal 48-Hour Definitive Test
EPA-821-R-02-012: Section 9 Method 2007

**CorrLine International – CorrX
Run-off from treated metal plate**

Test Organisms Age: 5 Days Old Test Organisms Source: EE

Test Initiation At: (A) on 9/13 /13

Counted by: Carli Young QC/QA by: _____
Loaded by: _____ Organism Lot # mb-117013

Exposure Chamber: 16 oz. plastic cups. Feeding: None.

Survival Data

Treatment % CTS													
Time	REP	LPC White	REP	0.10 Blue	REP	1.00 Green	REP	2.00 Yellow	REP	4.00 Red	REP	8.00 Black	Initials
0 HR	1	8	3	8	5	8	7	8	9	8	11	8	9/3/13
	2	8	4	8	6	8	8	8	10	8	12	8	
	(A)	///		///		///		///		///		///	(A)
24 HR 1014	1	8	3	8	5	8	7	8	9	8	11	8	9/4/13
	2	8	4	8	6	8	8	8	10	8	12	8	
		///		///		///		///		///		///	JA
48 HR 1327	1	8	3	8	5	8	7	8	9	8	11	8	9/5/13
	2	8	4	8	6	8	8	8	10	8	12	8	
		///		///		///		///		///		///	MR
72 HR 1341	1	8	3	8	5	8	7	8	9	7	11	4	9/6/13
	2	7	4	8	6	8	8	8	10	4	12	0	
		///		///		///		///		///		///	JA
96 HR 1154	1	8	3	8	5	8	7	8	9	3	11	1	9/7/13
	2	7	4	8	6	8	8	8	10	2	12	0	
		///		///		///		///		///		///	ME
% Surv	93.8		100		100		100		31.337.5(A)		4.3		

Data Entry by: Veronica Mc New
Double Data Entry: Veronica Mc New or
QA/QC Officer: Alan H. O'Neil

CorrX, Run-off from treated metal plate

Wrong data 09/13/13 Q-5821-13
NOEC/LOEC; LC50

(A) Data not recorded
09/10/13 ME

M. bahia Water Quality Data

All Treatments: Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

0 HR		Treatment % CTS						
9/3/13		LPC	0.10	1.00	2.00	4.00	8.00	Meter #
DO	I	7.1	7.2	7.2	7.0	7.1	7.0	S7
Temp	I	24.5	24.5	24.5	24.4	24.0	24.5	1B
Salinity	I	24.9	25.0	24.8	24.5	24.0	23.2	1B
Tech Initials: CMB		Time: 1330						

Comments _____

24 HR		Treatment % CTS						
9/4/13		LPC	0.10	1.00	2.00	4.00	8.00	Meter #
DO	F	6.7	6.6	5.5	4.9	3.8	2.6	S7
Temp	F	26.0	25.9	25.9	25.8	25.9	25.9	1B
Salinity	F	24.9	25.2	25.0	24.7	24.2	23.4	1B
pH	F	8.0	7.9	7.8	7.5	7.4	7.2	A93
Tech Initials: CMB		Time: 0849						

Comments began aeration
 at 34 ml per min
 on DAY 4 9-4-13
 JA

48 HR		Treatment % CTS						
9/5/13		LPC	0.10	1.00	2.00	4.00	8.00	Meter #
DO	F	6.6	6.8	7.0	6.9	6.9	6.8	S7
Temp	F	24.9	25.0	25.1	25.1	25.0	25.0	1B
Salinity	F	25.2	25.3	25.3	24.9	24.5	23.8	1B
pH	F	7.9	8.0	8.0	8.0	8.0	8.0	A93
Tech Initials: CMB		Time: 0834						

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

M. bahia Water Quality Data

All Treatments: Temp., 23.5 to 26.4°C. Initial & Final Dissolved Oxygen (DO): 4.0 to 7.5 mg/l.
 LPC: Initial Salinity, 24.5 to 25.4 ppt. I: initial water quality. F: final water quality.

48 HR		Treatment % CTS						
9/5/13		LPC	0.10	1.00	2.00	4.00	8.00	Meter #
DO	I	7.0	7.0	7.0	7.0	7.0	7.1	S7
Temp	I	24.5	24.5	24.5	24.5	24.5	24.5	1B
Salinity	I	24.5	24.9	24.8	24.5	24.1	23.1	1B
Tech Initials:		Cy			Time:			0958

Comments _____

72 HR		Treatment % CTS						
9/6/13		LPC	0.10	1.00	2.00	4.00	8.00	Meter #
DO	F	6.9	7.0	7.0	7.0	7.0	6.9	S7
Temp	F	24.9	24.9	25.1	25.0	24.8	25.0	1B
Salinity	F	24.8	25.2	25.0	24.9	24.6	23.4	1B
pH	F	7.9	7.9	8.0	8.0	7.9	7.9	A93
Tech Initials:		Cy			Time:			0841

Comments _____

96 HR		Treatment % CTS						
9/7/13		LPC	0.10	1.00	2.00	4.00	8.00	Meter #
DO	F	6.3	6.4	6.5	6.7	6.7	6.5	S7
Temp	F	25.2	25.3	25.4	25.4	25.5	25.6	1B
Salinity	F	25.1	25.7	25.9	25.3	24.3	23.5	1B
pH	F	7.8	7.8	7.9	8.0	8.0	7.9	A93
Tech Initials:		CM MK			Time:			0739

Comments _____

DO: mg/l pH: su Salinity: ppt Temp: °C

Acute Toxicity Test-96 Hr Survival

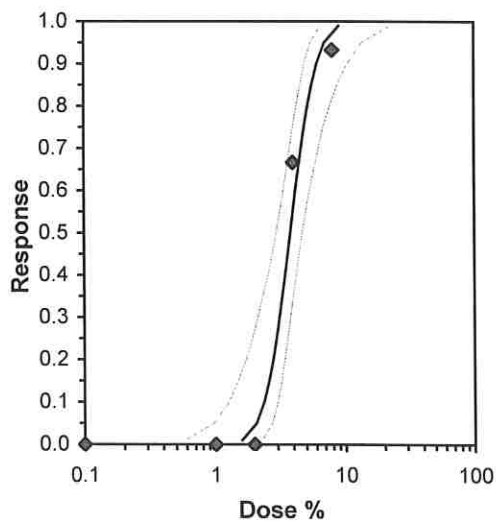
Start Date: 9/3/2013	Test ID: mb582113	Sample ID: LAB-Lab Sample
End Date: 9/7/2013	Lab ID: EE-Environmental Enterprise	Sample Type: PRD-Product
Sample Date:	Protocol: EPAM 02-EPA Marine	Test Species: MY-Mysidopsis bahia
Comments:		

Conc-%	1	2
PC-LP Control	1.0000	0.8750
0.1	1.0000	1.0000
1	1.0000	1.0000
2	1.0000	1.0000
4	0.3750	0.2500
8	0.1250	0.0000

Conc-%	Mean	N-Mean	Transform: Untransformed					N	Number Resp	Total Number
			Mean	Min	Max	CV%				
PC-LP Control	0.9375	1.0000	0.9375	0.8750	1.0000	9.428	2	1	16	
0.1	1.0000	1.0667	1.0000	1.0000	1.0000	0.000	2	0	16	
1	1.0000	1.0667	1.0000	1.0000	1.0000	0.000	2	0	16	
2	1.0000	1.0667	1.0000	1.0000	1.0000	0.000	2	0	16	
4	0.3125	0.3333	0.3125	0.2500	0.3750	28.284	2	11	16	
8	0.0625	0.0667	0.0625	0.0000	0.1250	141.421	2	15	16	

Auxiliary Tests	Statistic	Critical	Skew	Kurt
Normality of the data set cannot be confirmed				
Equality of variance cannot be confirmed				

Parameter	Value	SE	95% Fiducial Limits		Maximum Likelihood-Probit						
			Control	Chi-Sq	Critical	P-value	Mu	Sigma	Iter		
Slope	6.1687	1.61618	3.00098	9.33642	0.0625	3.7048	7.81473	0.29516	0.57873	0.16211	11
Intercept	1.42999	0.99971	-0.5295	3.38943							
TSCR	0.01635	0.01821	-0.0193	0.05204							
Point	Probits	%	95% Fiducial Limits								
EC01	2.674	1.59078	0.57785	2.26299							
EC05	3.355	2.05156	0.96144	2.71429							
EC10	3.718	2.34952	1.25567	3.00384							
EC15	3.964	2.57463	1.49888	3.2264							
EC20	4.158	2.76883	1.72075	3.42413							
EC25	4.326	2.94706	1.93199	3.6129							
EC40	4.747	3.44874	2.54316	4.2066							
EC50	5.000	3.79079	2.94753	4.69249							
EC60	5.253	4.16678	3.35552	5.32916							
EC75	5.674	4.87608	4.00447	6.8441							
EC80	5.842	5.18997	4.25159	7.63666							
EC85	6.036	5.58143	4.53637	8.72024							
EC90	6.282	6.11619	4.89531	10.3607							
EC95	6.645	7.00447	5.44099	13.473							
EC99	7.326	9.03341	6.55239	22.3269							



ENVIRONMENTAL ENTERPRISES USA, INC.

58485 Pearl Acres Rd., Suite D

Slidell, Louisiana 70461

(985) 646-2787

Kit No. 480A NPS

CHAIN - OF - CUSTODY RECORD

Client: CorrLine International

Contact Person: Barbara Tompkins-Brown

Spec

Address: 8 East Greenway Plaza

Phone#: 713 625 7519

Suite 910

P.O. #

Houston, Texas, 77046

Email: btompkins-brown@corrline.com

Fedex # 8995 9499 7160

Project:

Product Sample Description	Date Collected	Time Collected	No. of Containers	Analysis Request	S/R No.
Corr X Runoff from treated metal plate	8/27	6:15	1	M. bahia 96-hr Acute RFT, Method 2007.0 & beryllina 96-hr Acute RFT, Method 2006.0 M. bahia 7-day Chronic, Method 1007.0 & beryllina 7-day Chronic, Method 1006.0	Q-582-13 Q-525-13 Q-625-13
					used sample received 09/20/13

Collected By: Kirk Chrisman
 Received By: Nicole Bailey
 Received By: Vanessa Holt-Ven
 Date & Time: 8/27 6:20
 Date & Time: 8/28/13 0915
 Date & Time: 08/28/13 0920
 Relinquished By: Nicole Bailey
 Relinquished By: Vanessa Holt-Ven
 Date & Time: 8/28/13 0919
 Date & Time: 10/14/13

REMAILED
10/14/13

1232
1500

FedEx NEW Package Express
US Airbill

FedEx Tracking Number

8995 9499 7160

1 From This portion may be removed for Recipient's records
Date 2/27/13 FedEx Tracking Number 15899594997160
Sender's Name CorrLine Int'l, LLC Phone 281 633-9335
Company Mike Chrisman
Address 8 Gleerway Plaza #910
City Houston State TX ZIP 77046
Darr./Room/Sub/Rm

RECIPIENT: PML HERE

2 Your Internal Billing Reference

3 To Recipient's SHIPPING AND RECEIVING Phone 955 648-2787

Company ENVIRONMENTAL ENTERPRISES USA

Address 5885 PEARL ACRES RD STE D
We cannot deliver to P.O. boxes or P.O. ZIP codes.

Address Use this line for the HOLD location address or for combination of your shipping address.

City SLIDELL State LA ZIP 70451-5400

0100055469



8995 9499 7160

MUR4
0215
0215
0215

4 Express Package Service *To most locations. For per package fee, see the new FedEx Express Freight US Airbill.

Next Business Day
FedEx First Overnight
FedEx 2Day
FedEx Standard Overnight
FedEx Express Saver

5 Packaging *Declared value limit \$500.
FedEx Envelope*
FedEx Pak*
FedEx Tube
Other

6 Special Handling and Delivery Signature Options
SATURDAY Delivery
Indirect Signature
Direct Signature

7 Payment Bill for:
Sender Station
Recipient
Third Party
Credit Card
Cash/Check
Obtain recip. Acct. No.

Total Packages 4
Total Weight 24 lbs.

0100055469

8995 9499 7160

ENVIRONMENTAL ENTERPRISES USA, INC.
SAMPLE RECEIPT / ACCEPTANCE (SRA) FORM

CLIENT: Cool Line
 DATE RECEIVED: 9-20-13
 LOCATION: NA

KIT NO. 1581A
 CL NO. P808 LAB NO. _____

NA

SAMPLE RECEIPT:

- Sample Kit Supplied by: EE USA. , Client....., ???....., Other.....
 Ice Chest. , Cardboard Box....., Styrofoam Box....., Other..... How many containers in kit? 2
- Ice chest received... Circle one; *delivered by Hot Shot, FEDEX, UPS, Client, etc. mark NA.
 *NA or SB: Fridge, Ice & H₂O, [Dry, H₂O, Ice packs, Other, (Temp.....°C, Temp ID#.....)]
 At EE USA: Ice & H₂O, Dry, H₂O, Ice packs (Frozen? Yes....or No.....), Other.....
 If Ice & H₂O received... How? Loose, Bagged, Bottled, or Other... Comment: FEX 598 9499 742
- Sample container(s) in good condition (sealed & unbroken)? YES. NO.....
- Sample container label(s) filled out completely? YES..... NO. N/A.....
 If not, mark all that apply. For O&G (PW) OCSG & Well #'s N/A. For O&G (WF) OCSG N/A
 a) Date & time collected..... c) OCSG number..... _____
 b) Collected by..... _____ d) Well number..... _____
- Chain-of-Custody form (COC) filled out completely? YES..... NO.
 If not, mark all that apply.
 a) No COC..... f) Date & time collected..... _____
 b) Collected by..... _____ g) Received by..... _____
 c) Relinquished by..... _____ h) Date and/or time of transfer... _____
 d) Location..... _____ i) Waste type..... _____
 e) Company name..... _____
- Custody seal(s) received with this sample kit? YES..... NO.
 Were custody seals used? YES..... NO..... And if used, were they intact? YES.....NO.....

COMMENTS:

Information recorded by: J. DiStella 9/20/13

SAMPLE ACCEPTANCE:

TOX: EFF ___ CTS ___ PW ___ DF ___; ANALY ___ BIOD ___;
 O&G: PW ___ WF ___; PROD.: NCP ___ WAF ___ SBF ___;
 ADD.: DF ___ CTS ___; OTHER: _____

- Was each sample container appropriate (EPA Protocol)? YES..... NO.....
 Plastic..... Glass..... Number of samples for location?.....
- Does the recorded information on the COC and label agree? YES..... NO.....
 Client Sample ID, Collection location, date, & time. Collected by.
- Was sufficient amount of each sample received? YES..... NO.....
 Container size....., Estimated Volume....., Head space..... (mls or liters).
- Was each sample received within the proper holding time? YES..... NO.....
- Was each sample received at the proper temperature? (See COC for temp) YES..... NO.....

Oil & Grease Lab Only:

- Sample verified for proper acid preservation & temp within 1 hour of sample receipt? YES..... NO.....
- Is the initial pH <2 su? YES..... NO.....
 If no, how many mls of 6NHCL was added to make pH <2 su?mls.....OL#

COMMENTS & CLIENT CONTACT INFO (name, date, instructions):

Information recorded by: _____ / ___ /13

NA=not applicable, SB=Shorebase, or CL=Certified Lot Number, PW= Produced Water, WF=Well Fluid

KAH # 158A TO

1500

FedEx Express **NEW Package**
US Airbill

FedEx Tracking Number
8995 9499 7491

1 From **This portion can be removed for Recipient's records.**
Date **9/19/03** FedEx Tracking Number **899594997491**

Sender's Name **Kirk Chrisman** Phone
Company **Carlisle TX 41**
Address **8 Glenway Blvd. #910**
City **Houston** State **TX** ZIP **77066**

2 Your Internal Billing Reference

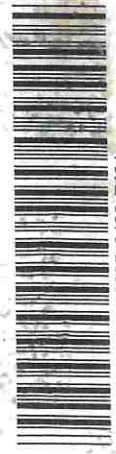
3 To Recipient's Name **SHIPPING AND RECEIVING** Phone **985 646-2787**

Company **ENVIRONMENTAL ENTERPRISES USA**

Address **58485 PEARL ACRES RD STE 5**
We cannot deliver to P.O. boxes or P.O. ZIP codes.

Address **SLIDELL** State **LA** ZIP **70461** - 3400

City **SLIDELL** State **LA** ZIP **70461** - 3400



8995 9499 7491

MUR4
Form ID No. **0215**
Recipient's Copy

4 Express Package Service
NOTE: Service under has changed. Please select carefully.
* To most locations.

Next Business Day
 FedEx First Overnight
Earliest next business day delivery to select locations. Friday shipments will be delivered on Monday unless SATURDAY Delivery is selected.
 FedEx Priority Overnight
High business priority. Friday shipments will be delivered on Monday unless SATURDAY Delivery is selected.
 FedEx Standard Overnight
Next business afternoon.
Saturday Delivery NOT available.

5 Packaging *Declared value limit \$500.
 FedEx Envelope* FedEx Pak* FedEx Box FedEx Tube Other

6 Special Handling and Delivery Signature Options
 SATURDAY Delivery
NOT available for FedEx Standard Overnight, FedEx 2Day A.M., or FedEx Express Saver.

No Signature Required
Package may be left without obtaining a signature for delivery.
 Direct Signature
Someone at recipient's address may sign for delivery. Fee applies.
 Indirect Signature
Indirect signature available at recipient's address may sign for delivery. Fee applies.
Does this shipment contain dangerous goods?
 No Yes
As per attached Shipper's Declaration not required. Dry Ice (Dry Ice 2, UN 1845) Cargo Aircraft Only

7 Payment Bill to:
 Sender (You bill) Recipient (You bill)
Enter FedEx Acct. No. or Credit Card No. below. Obtain receipt Acct. No. Credit Card Cash/Check

Total Packages **2** Total Weight **1.7** lbs.
Total Packages Total Weight
*Weight is limited to 50 lbs unless you declare a higher value. See the current FedEx Saver® Guide for details.

ENVIRONMENTAL ENTERPRISES USA, INC.
SAMPLE RECEIPT / ACCEPTANCE (SRA) FORM

CLIENT: ComLine
 DATE RECEIVED: 9-20-13
 LOCATION: 1/A

KIT NO. 157A
 CL NO. 1808 LAB NO. _____
1808 Q-626-13
Q-625-13

SAMPLE RECEIPT:

- Sample Kit Supplied by: EE USA....., Client....., ???....., Other.....
 Ice Chest....., Cardboard Box....., Styrofoam Box....., Other..... How many containers in kit? 2
- Ice chest received... Circle one; *delivered by Hot Shot, FEDEX, UPS, Client, etc. mark NA.
 *NA or SB: Fridge, Ice & H₂O, [Dry, H₂O, Ice packs, Other, (Temp.....°C, Temp ID#.....)]
 At EE USA: Ice & H₂O, Dry, (H₂O), Ice packs (Frozen? Yes....or No....), Other.....
 If Ice & H₂O received... How? Loose, Bagged, Bottled, or Other... Comment: 899594997480
- Sample container(s) in good condition (sealed & unbroken)? YES... NO.....
- Sample container label(s) filled out completely? YES..... NO... N/A.....
 If not, mark all that apply. For O&G (PW) OCSG & Well #'s N/A. For O&G (WF) OCSG N/A
 a) Date & time collected..... ✓ c) OCSG number.....
 b) Collected by..... d) Well number.....
- Chain-of-Custody form (COC) filled out completely? YES..... NO...
 If not, mark all that apply.
 a) No COC..... f) Date & time collected.....
 b) Collected by..... g) Received by.....
 c) Relinquished by..... h) Date and/or time of transfer...
 d) Location..... i) Waste type.....
 e) Company name.....
- Custody seal(s) received with this sample kit? YES..... NO...
 Were custody seals used? YES..... NO..... And if used, were they intact? YES.....NO.....

COMMENTS:

Information recorded by: [Signature] 9/20/13

SAMPLE ACCEPTANCE: TOX: EFF ___ CTS ___ PW ___ DF ___; ANALY ___ BIOD ___;
 O&G: PW ___ WF ___; PROD.: NCP ___ WAF ___ SBF ___;
 ADD.: DF ___ CTS ___; OTHER: product

- Was each sample container appropriate (EPA Protocol)? YES... NO.....
 Plastic... Glass..... Number of samples for location? 2
- Does the recorded information on the COC and label agree? YES... NO.....
 Client Sample ID, Collection location, date, & time. Collected by.
- Was sufficient amount of each sample received? YES... NO.....
 Container size 4.20, Estimated Volume 80, Head space 0 (mls or liters).
- Was each sample received within the proper holding time? YES..... NO..... N/A
- Was each sample received at the proper temperature? (See COC for temp) YES..... NO..... N/A

Oil & Grease Lab Only:

- Sample verified for proper acid preservation & temp within 1 hour of sample receipt? YES..... NO.....
- Is the initial pH <2 su? YES..... NO.....
 If no, how many mls of 6NHCL was added to make pH <2 su?mls.....OL#

COMMENTS & CLIENT CONTACT INFO (name, date, instructions):

Information recorded by: 1/2 09/20/13

NA=not applicable, SB=Shorebase, or CL=Certified Lot Number, PW= Produced Water, WF=Well Fluid

ENVIRONMENTAL ENTERPRISES USA, INC.
SAMPLE RECEIPT / ACCEPTANCE (SRA) FORM

CLIENT: Corrine
 DATE RECEIVED: 8/28/13
 LOCATION: Corrx

KIT NO. 480A
 CL NO. P811 LAB NO. _____

SAMPLE RECEIPT:

- Sample Kit Supplied by: EE USA....., Client....., ???....., Other.....
 Ice Chest....., Cardboard Box....., Styrofoam Box....., Other..... How many containers in kit? 1.....
- Ice chest received... Circle one; *delivered by Hot Shot, FEDEX, UPS, Client, etc. mark NA.
 *NA or SB: Fridge, Ice & H₂O, [Dry, H₂O, Ice packs, Other, (Temp.....°C, Temp ID#.....)]
 At EE USA: Ice & H₂O, Dry, H₂O, Ice packs (Frozen? Yes.....or No.....), Other.....
 If Ice & H₂O received... How? Loose, Bagged, Bottled, or Other...Comment: _____
- Sample container(s) in good condition (sealed & unbroken)? YES..... NO.....
- Sample container label(s) filled out completely? YES..... NO..... N/A.....
 If not, mark all that apply. For O&G (PW) OCSG & Well #'s N/A. For O&G (WF) OCSG N/A
 a) Date & time collected..... c) OCSG number.....
 b) Collected by..... d) Well number.....
- Chain-of-Custody form (COC) filled out completely? YES..... NO.....
 If not, mark all that apply. *wrong data NPB 8/28/13*
 a) No COC..... f) Date & time collected.....
 b) Collected by..... g) Received by.....
 c) Relinquished by..... h) Date and/or time of transfer...
 d) Location..... i) Waste type.....
 e) Company name.....
- Custody seal(s) received with this sample kit? YES..... NO.....
 Were custody seals used? YES..... NO..... And if used, were they intact? YES.....NO.....

COMMENTS: (4) Blank

Information recorded by: NPB 8/28/13

SAMPLE ACCEPTANCE:

TOX: EFF__ CTS__ PW__ DF__ ; ANALY__ BIOD__ ;
 O&G: PW__ WF__ ; PROD.: NCP__ WAF__ SBF__ ;
 ADD.: DF__ CTS__ ; OTHER: _____

- Was each sample container appropriate (EPA Protocol)? YES..... NO.....
 Plastic..... Glass..... Number of samples for location?.....
- Does the recorded information on the COC and label agree? YES..... NO.....
 Client Sample ID, Collection location, date, & time. Collected by.
- Was sufficient amount of each sample received? YES..... NO.....
 Container size....., Estimated Volume....., Head space..... (mls or liters).
- Was each sample received within the proper holding time? YES..... NO.....
- Was each sample received at the proper temperature? (See COC for temp) YES..... NO.....

Oil & Grease Lab Only:

- Sample verified for proper acid preservation & temp within 1 hour of sample receipt? YES..... NO.....
- Is the initial pH <2 su? YES..... NO.....
 If no, how many mls of 6NHCL was added to make pH <2 su?mls..... OL#

COMMENTS & CLIENT CONTACT INFO (name, date, instructions):

Information recorded by: _____ / ____ / 13

NA=not applicable, SB=Shorebase, or CL=Certified Lot Number, PW= Produced Water, WF=Well Fluid

MUR-4

FedEx **NEW Package**
Express **US Airbill**

FedEx Tracking Number **8995 9499 7160**

1 From **This portion can be removed for Recipient's records.**
Date **8/27/13** FedEx Tracking Number **15-899594997160**

Sender's Name **Corline Int'l, LLC** Phone **281 635-9335**
Company **Kirk Chrisman**
Address **8 Greenway Plaza #910**
City **Houston** State **Tx** ZIP **77046**

Direct Floor/Suite/Room

2 Your Internal Billing Reference

3 To Recipient's Name **SHIPPING AND RECEIVING** Phone **985 646-2787**

Company **ENVIRONMENTAL ENTERPRISES USA**

Address **58485 PEARL ACRES RD STE D**
We cannot deliver to P.O. boxes or P.O. ZIP codes.

Dept./Floor/Suite/Room

Address **SLIDELL** State **LA** ZIP **70461-5400**

Use this line for the HOLD location address or for continuation of your shipping address.

City **SLIDELL**



8995 9499 7160

Recipient's Copy

From ID No. **0215**

4 Express Package Service **2 or 3 Business Days**
NOTE: Service order has changed. Please select carefully.

Next Business Day
 FedEx First Overnight
Earliest next business morning delivery to select locations. Friday shipments will be delivered on Monday unless SATURDAY Delivery is selected.
 FedEx Priority Overnight
Next business morning. Friday shipments will be delivered on Monday unless SATURDAY Delivery is selected.
 FedEx Standard Overnight
Next business afternoon. Sunday Delivery NOT available.

5 Packaging *Declared value limit \$500.
 FedEx Envelope* FedEx Pak* FedEx Box FedEx Tube Other

6 Special Handling and Delivery Signature Options

SATURDAY Delivery
NOT available for FedEx Standard Overnight, FedEx 2D or A.M., or FedEx Express Saver.
 No Signature Required
Package may be delivered without obtaining a signature for delivery.
 Direct Signature
Shipper's signature required. Address may sign for delivery. Fee applies.
 Indirect Signature
Signature of addressee or recipient's address may sign for delivery. For residential deliveries only. Fee applies.

Does this shipment contain dangerous goods?
 No Yes
One box must be checked. Shipper's Declaration not required.
 Dry Ice
Dry Ice 2, UN 1845 x kg
 Cargo Aircraft Only

7 Payment Bill to:
 Sender Recipient Third Party Credit Card Cash/Check

Total Packages **4** Total Weight **24** lbs.
Obtain recip. Acct. No. **633**

Your liability is limited to \$100 unless you declare a higher value. See the current FedEx Service Guide for details.

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